

## Variations of pump-probe spectroscopy

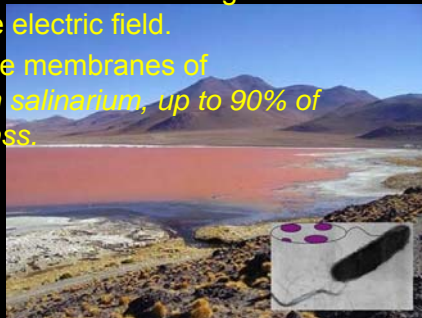
## Bacteriorhodopsin, summary of:

- Widely investigated:
  - Nature – 34 papers (1990 – 2012)
  - Science – 43 papers(1990 – 2012)
  - PNAS -173 papers (1990 – 2012)

(Source: Web of science)

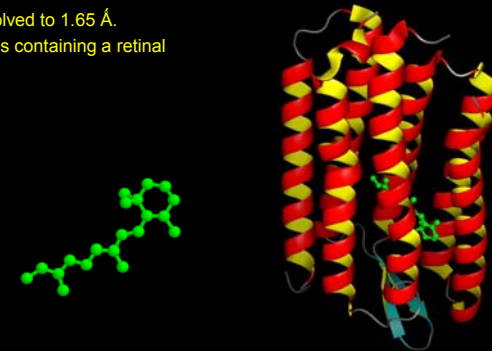
## Function:

- Light drive proton pump that pushes protons across the membrane against the direction of the electric field.
- Found in purple membranes of *Halobacterium salinarum*, up to 90% of membrane mass.



## Structure: membrane protein

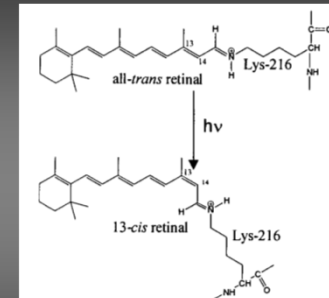
- Structure resolved to 1.65 Å.
- 7 alpha helices containing a retinal chromophore



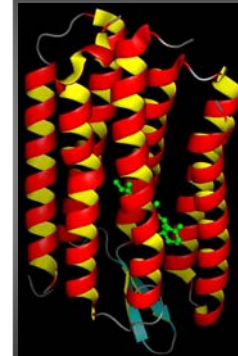
## Advantages:

- Chemically stable and photostable
- Well known structure, easy to crystalize
- Bacteria grow a lot of it
- Fast, photoactive and therefore interesting
- Can be used as biomolecular tool or a model system for photoreactions

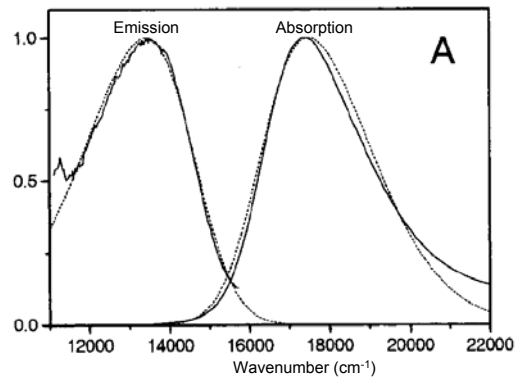
## Isomerization of Retinal



**Fig. 1.** Retinal with a protonated Schiff base. In bR, the retinal is housed within the hydrophobic interior of a seven-helix bundle and is covalently linked to Lys<sup>216</sup>. Upon absorbing a photon, the retinal isomerizes around the C<sub>13</sub>-C<sub>14</sub> bond.

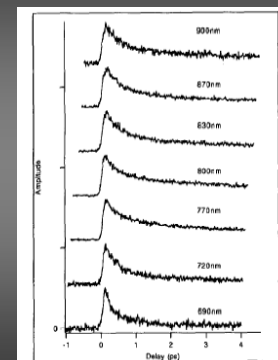


## Absorption and emission



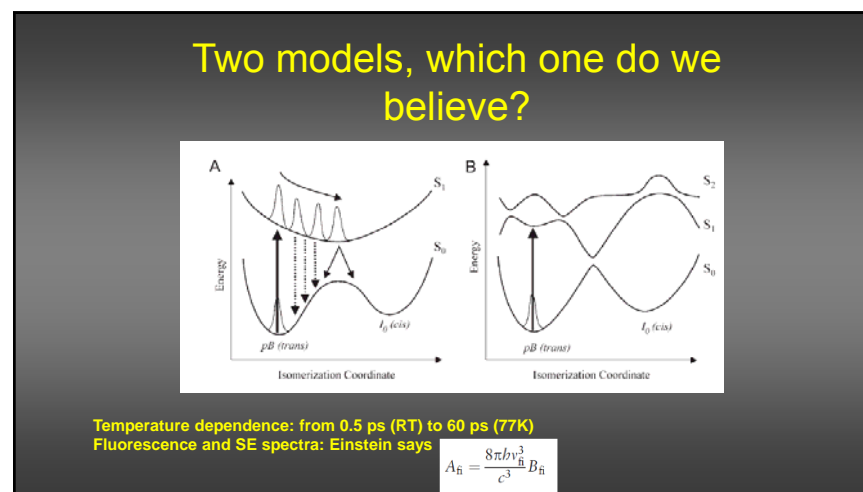
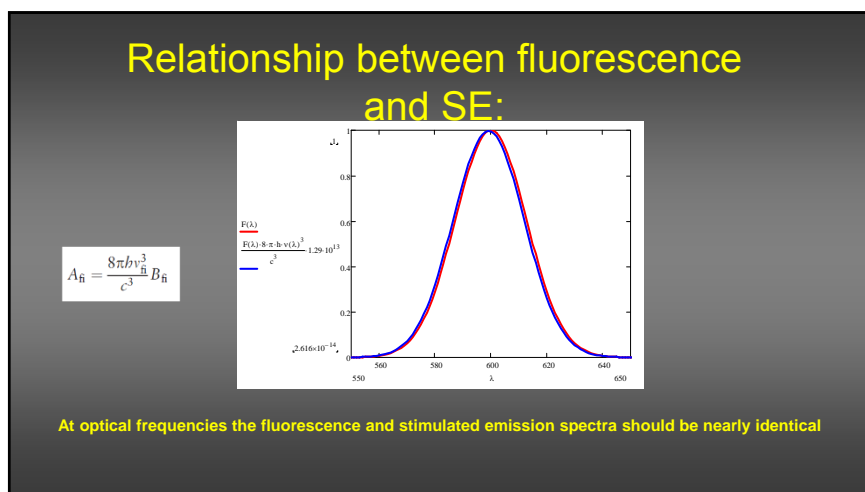
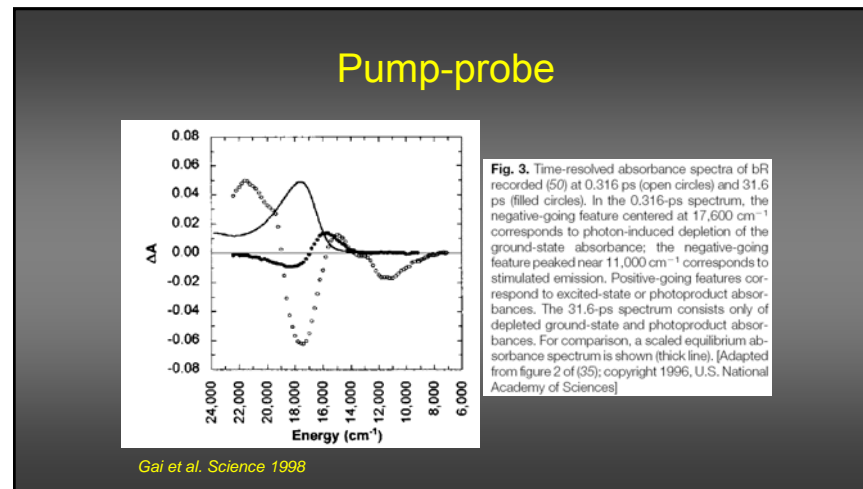
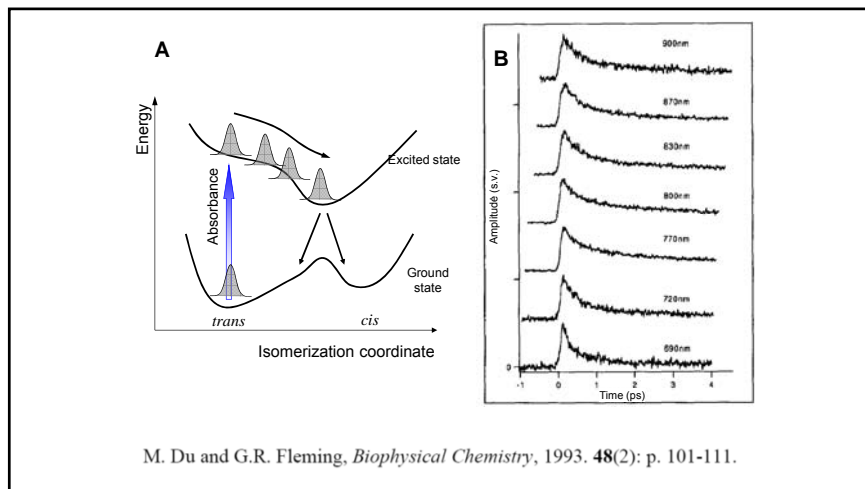
## Observation:

- The decays can be well fitted as a sum of three exponential decay components with time constants in the range of 90 fs-240 fs, 0.6 ps-0.9 ps, and 9.0-13.0 ps.
- Fluorescence disappears on sub-ps time scale!



**Fig. 3.** Fluorescence decays at different detection wavelengths measured over 5 ps scan length.

*Du et al. Biophysical chemistry, 1993*



No detailed information about the structural changes of the molecule

...solution: go infrared! ☺

The probe beam needs to be in the mid-IR

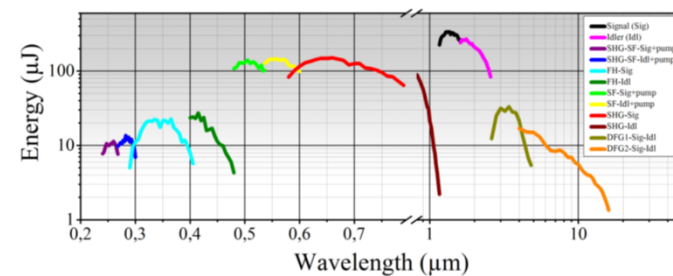
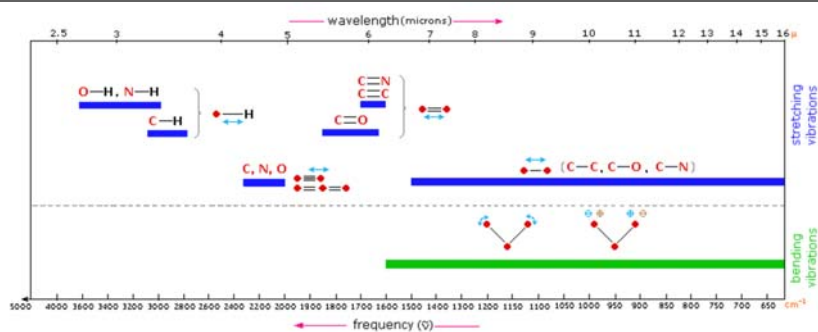


Fig. 11. Tuning curves of TOPAS-prime with VIS and IR extensions when pumped by the 55 fs, 1.75 mJ, 100 kHz burst-mode pulses from the PP-laser.



Molecular vibration frequencies



Vibrational features of *cis* appear within < 1ps!

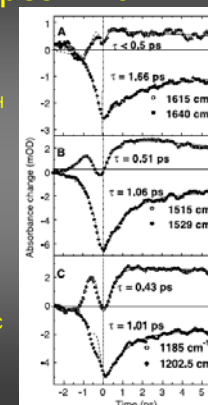
The IR spectrum of *cis* known from FTIR measurements forms in < 1 ps.

First evidence that it is isomerization that causes fluorescence to disappear so fast.

C—NH

C=C

C—C



Herbst et al. Science, 2002

## Comparison of vibrational features in the spectrum

Top: Visible pump – mid IR probe;  
Bottom: Step-scan FTIR (15 ns)

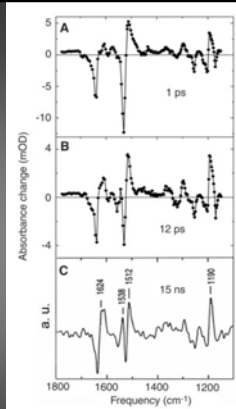


Fig. 2. IR difference spectra of bR with conditions as in Fig. 1 at (A) 1 ps and (B) 12 ps. (C) For comparison, a FTIR bR<sub>1010</sub>K difference spectrum at 15 ns after photoexcitation at room temperature [from (15)].

Herbst et al. Science, 2002

What is the function of the protein?...

....Solution: go UV! ☺

## Protein role in BR isomerization?...

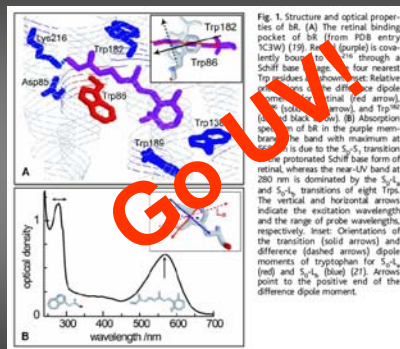
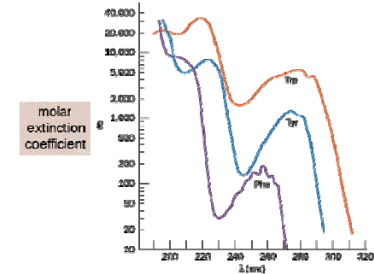
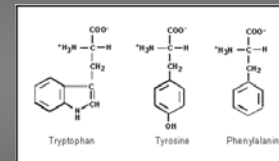


Fig. 1. Structure and optical properties of bR. (A) The retinal binding pocket of bR (from PDB entry 1C3W) (19). Ret (purple) is covalently bound to Lys50 through a Schiff base. The four nearest Trp residues (shown in red) relative to the difference dipole moment of the protonated Schiff base form of retinal (red arrow) are Trp216 (red arrow), Trp182 (red arrow), Trp180 (red arrow), and Trp90 (red arrow). (B) Absorption spectrum of bR in the purple membrane. The band with maximum at 280 nm is dominated by the  $S_0 \rightarrow S_1$  transition of the protonated Schiff base form of retinal, whereas the near-UV band at 260 nm is dominated by the  $S_0 \rightarrow S_1$  and  $S_0 \rightarrow S_2$  transitions of eight Trp. The vertical and horizontal arrows indicate the excitation wavelength and the range of probe wavelengths, respectively. Inset: Orientations of the transition (solid arrows) and difference (dashed arrows) dipole moments of tryptophan for  $S_0 \rightarrow S_1$  (red) and  $S_0 \rightarrow S_2$  (blue) (21). Arrows point to the positive end of the difference dipole moment.

Schenkl et al. Science 2005

## Protein role in BR isomerization?...



UV absorbance spectra of the three aromatic amino acids, phenylalanine, tryptophan, and tyrosine

## Protein reacting to electric field of retinal

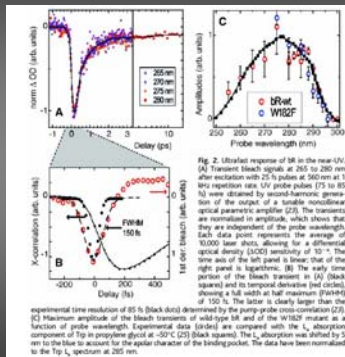
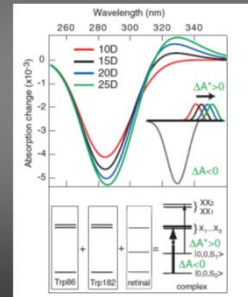


Fig. 2. Ultrafast response of IR in the near-UV. (A) Transient bleach signals at 265 to 285 nm after excitation with 25 fs pulses at 360 nm at 1 kHz repetition rate. UV probe pulses (75 to 85 fs) were obtained by second-harmonic generation of the output of a tunable monolithic optical parametric amplifier (OPA). The transients are normalized in amplitude, which shows that they are independent of the probe wavelength. Each data point represents the average of 10,000 laser shots, allowing for a differential optical density (DOD) sensitivity of  $10^{-4}$ . The time axis of the left panel is linear; that of the right panel is logarithmic. (B) The early time portion of the bleach transient in (A) (black squares) and its temporal derivative (red circles), showing a full width at half maximum (FWHM) of 150 fs. The factor is clearly larger than the experimental time resolution of 85 fs (black dots) determined by the pump-probe cross-correlation (27). (C) Maximum amplitude of the bleach transients of wild-type IR and of the W182F mutant as a function of probe wavelength. Experimental data (black squares) are compared with the  $L_{\alpha}$  absorption component of Trp in propylene glycol at  $-10^{\circ}\text{C}$  (25) (black squares). The  $L_{\alpha}$  absorption was shifted by 5 nm to the blue to account for the apolar character of the binding pocket. The data have been normalized to the Trp  $L_{\alpha}$  spectrum at 265 nm.

Schenkl et al. Science 2005

Tryptophan acts as an electric field probe inside the protein highlighting the electric field evolution after the excitation.



Complex:  
Retinal+2Trp's

## Protein reacting to electric field of retinal

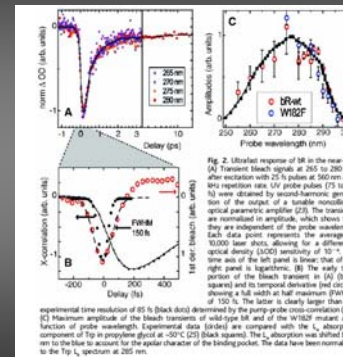
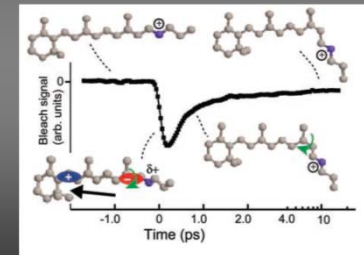


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Schenkl et al. Science 2005

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Adding a twist...

Advanced ultrafast spectroscopies

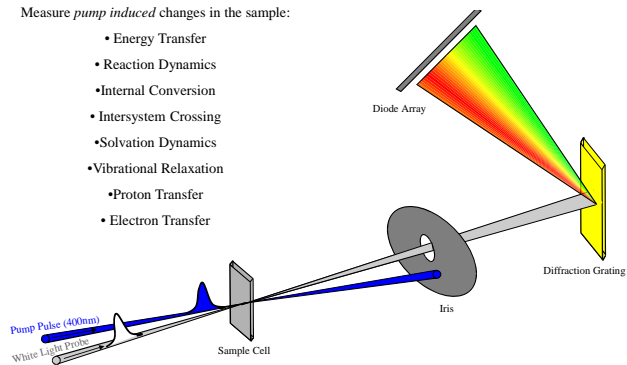
Twist #1:

Multi-pulse transient absorption

## Dispersed Pump-Probe Experimental Setup

Measure *pump induced* changes in the sample:

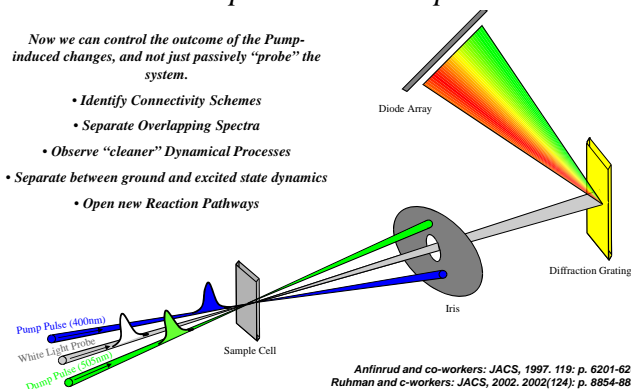
- Energy Transfer
- Reaction Dynamics
- Internal Conversion
- Intersystem Crossing
- Solvation Dynamics
- Vibrational Relaxation
- Proton Transfer
- Electron Transfer



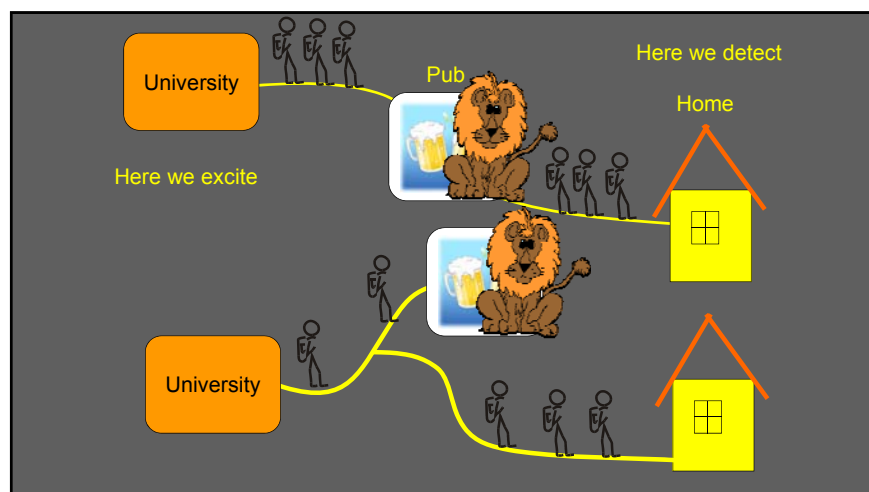
## Dispersed Multi-Pulse Experimental Setup

Now we can control the outcome of the Pump-induced changes, and not just passively "probe" the system.

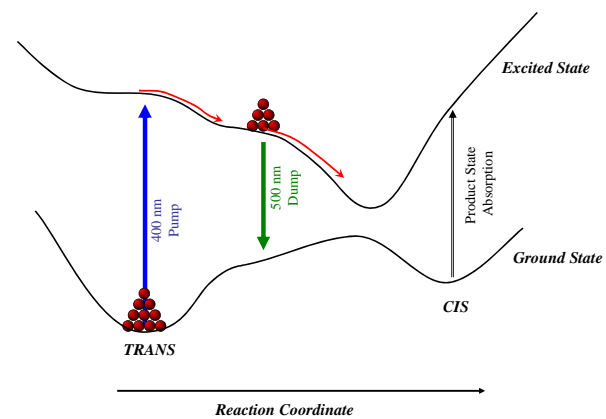
- Identify Connectivity Schemes
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- Observe "cleaner" Dynamical Processes
- Separate between ground and excited state dynamics
- Open new Reaction Pathways

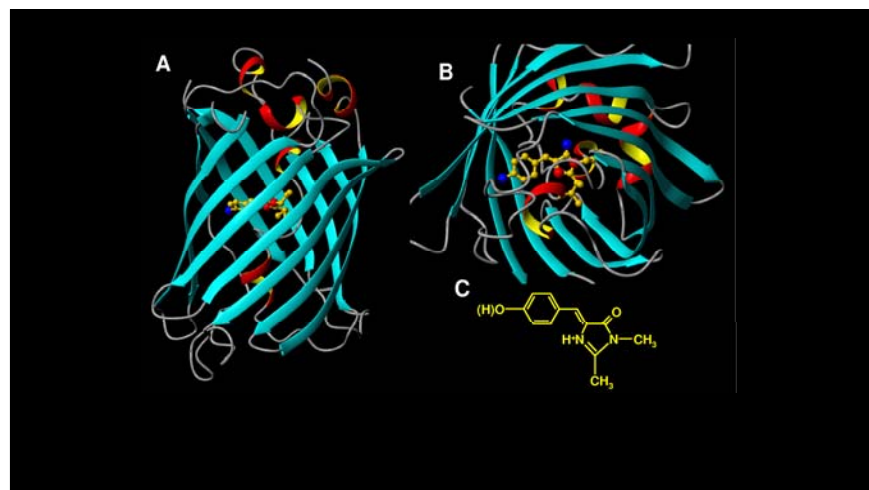
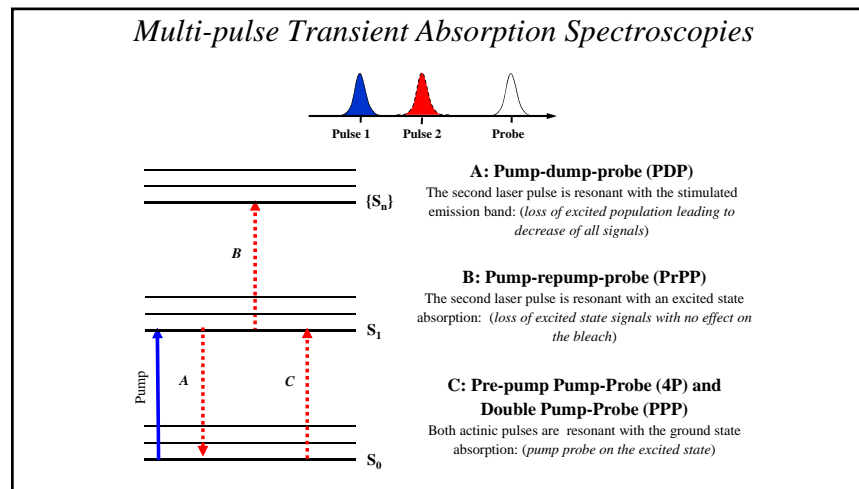
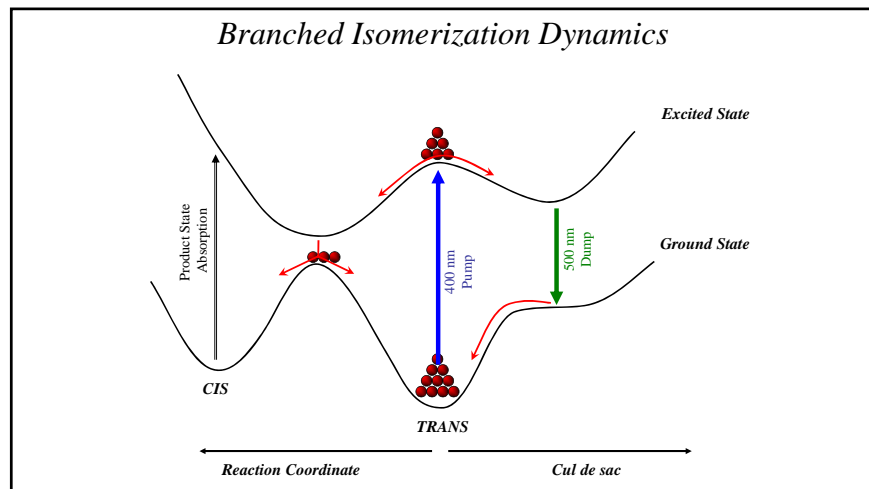


Anfinrud and co-workers: JACS, 1997, 119: p. 6201-6202  
 Ruhman and co-workers: JACS, 2002, 124: p. 8854-8858  
 Field and co-workers: Journal of Chemical Physics, 1988, 88(9): p. 5972-5974

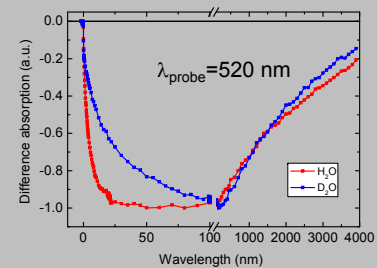


## Sequential Isomerization Dynamics



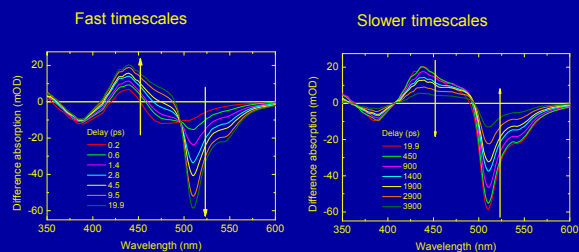


### Kinetic Isotope Effect: evidence for proton transfer

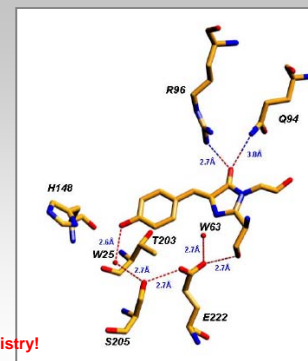
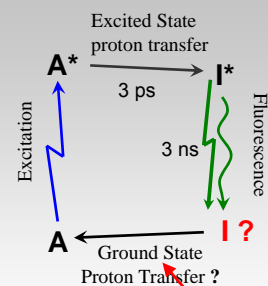




## Excited state proton transfer in GFP



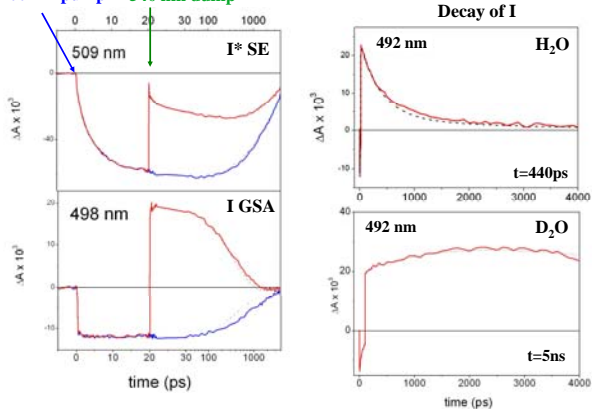
*Idea: try to dump the excited state!*



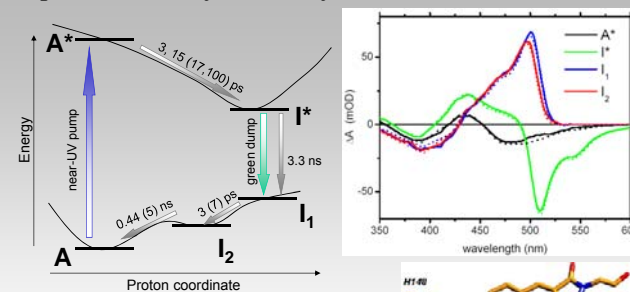
A fundamental reaction in biochemistry!

## Pump-dump-probe spectroscopy on GFP

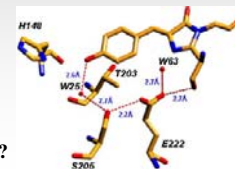
400 nm pump 540 nm dump



## Spectra and lifetimes of Hidden Intermediates

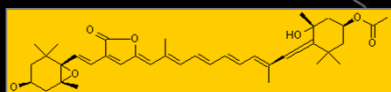


- I<sub>1</sub> max: 500 nm, I<sub>2</sub> max: 498 nm
- I<sub>1</sub> Stokes shift: 9 nm
- I<sub>1</sub> to I<sub>2</sub> evolution: H-bond rearrangement?

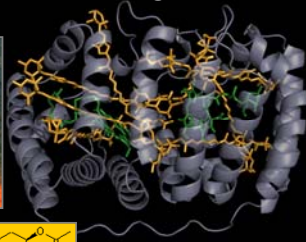


## Energy transfer in PCP: role of ICT state

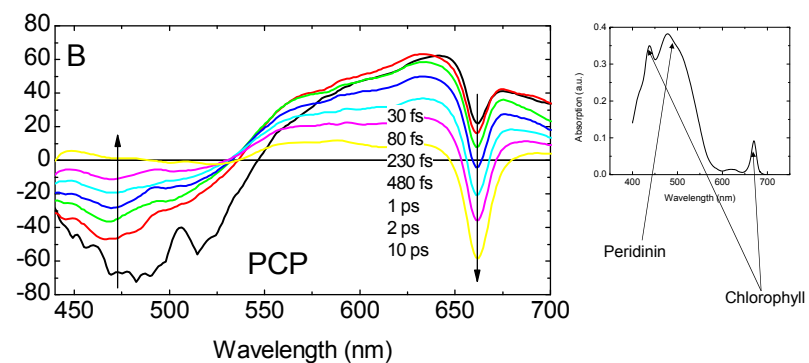
Red algae



PCP



## PCP harvesting light



## For warm up: peridinin in solution

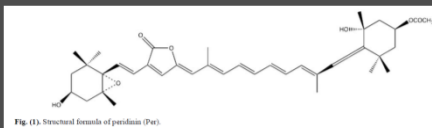


Fig. (1). Structural formula of peridinin (Per).

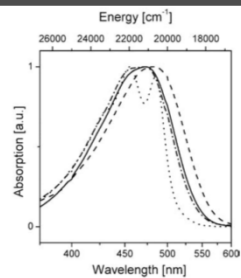
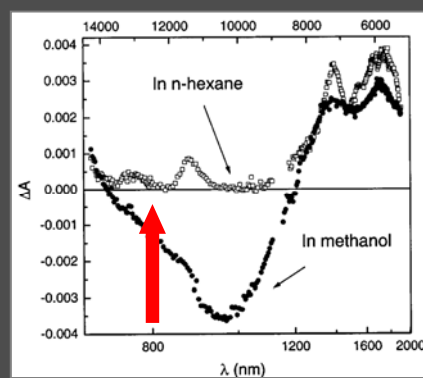


Fig. (2). Absorption spectra of Per in n-hexane (\*\*\*), acetonitrile (- · -), methanol (· · ·), and ethylene glycol (- - -). All spectra are normalized. Reprinted with permission from Zigmantas, D.; Hiller, R.G.; Yarsev, A.; Sundstrom, V.; Polivka, T. J. Phys. Chem. B 2003, 107, 5339-5348 Copyright 2003 American Chemical Society.

## Near-IR transient Absorption spectra of peridinin



Zigmantas *et al.*, JPC A 105, 2001

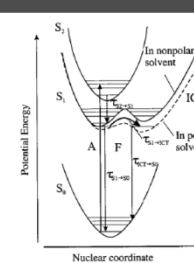
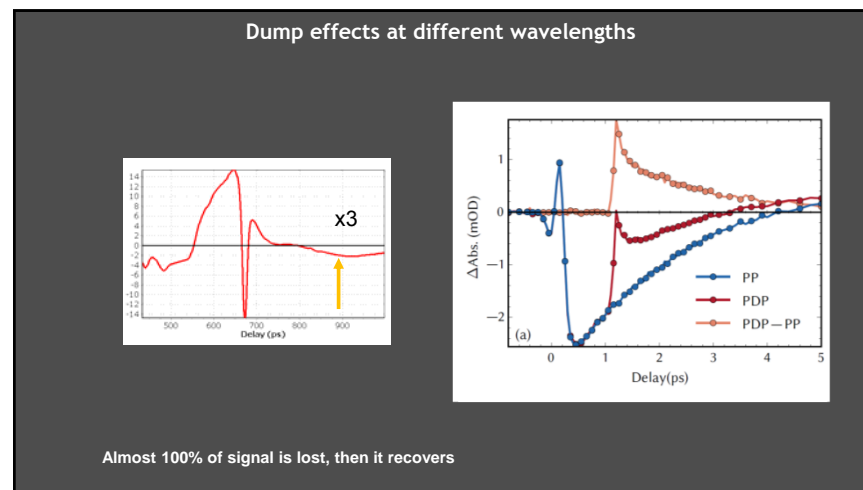
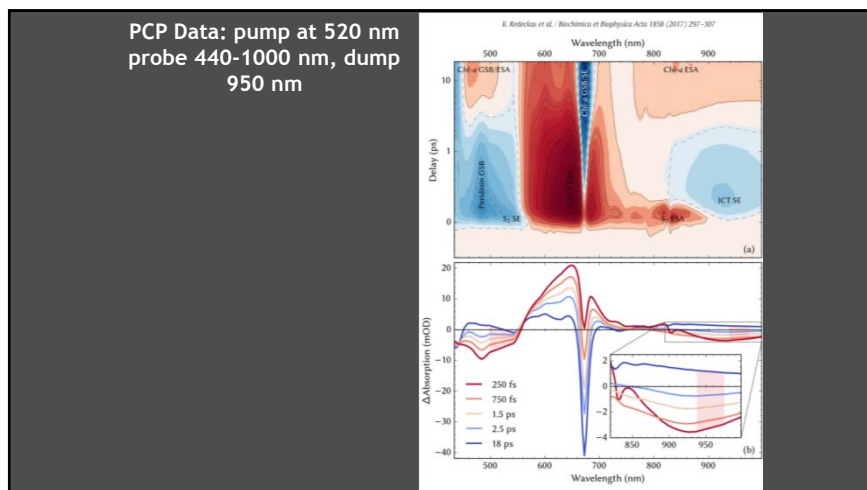
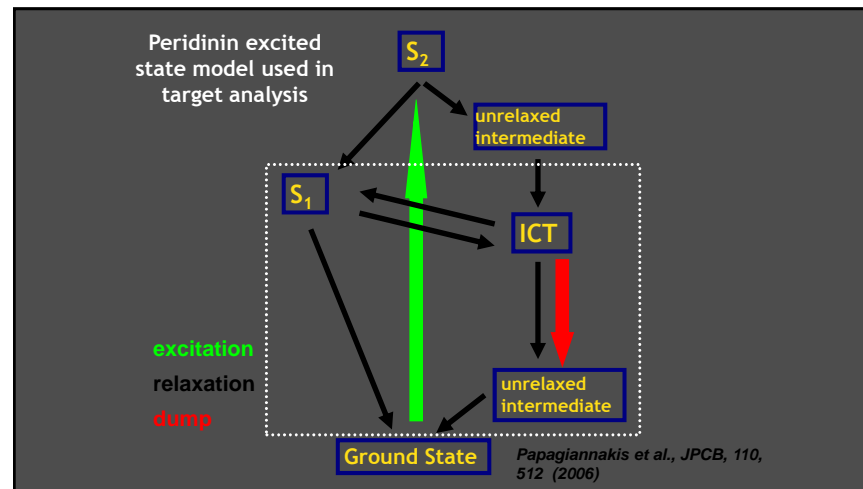
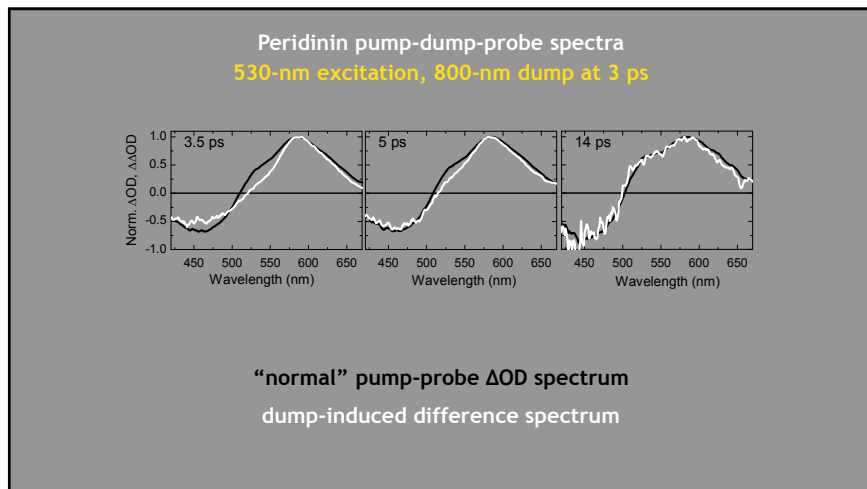
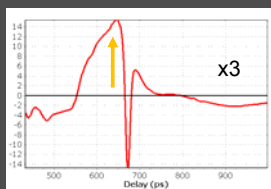


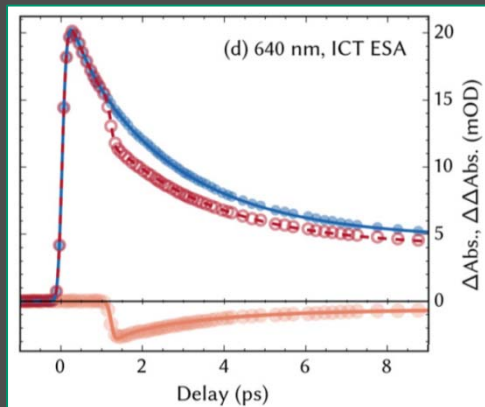
Figure 9. Potential energy surface diagram showing the ground, first, and second singlet excited states of peridinin in nonpolar and polar solvent environment. The S<sub>1</sub> state is strongly coupled with the ICT state. In nonpolar solvents, the barrier between the S<sub>2</sub> and ICT states is high and therefore the ICT state is not populated. In polar solvents, i.e., methanol, the polarity-affected barrier is diminished as the ICT state shifts to lower energy, therefore becoming populated. The transitions and corresponding time constants are shown for the peridinin in polar solvent.



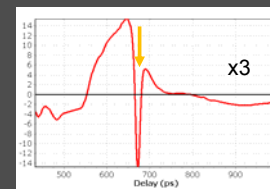
## Dump effects at different wavelengths



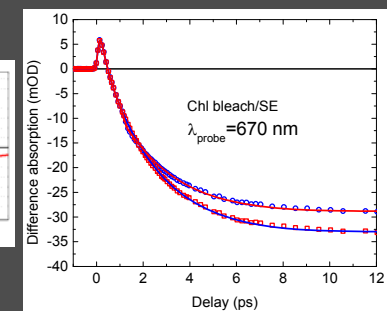
The loss is about 20% - overlapping signals?



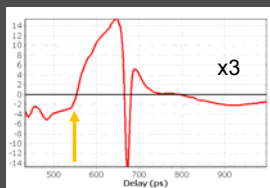
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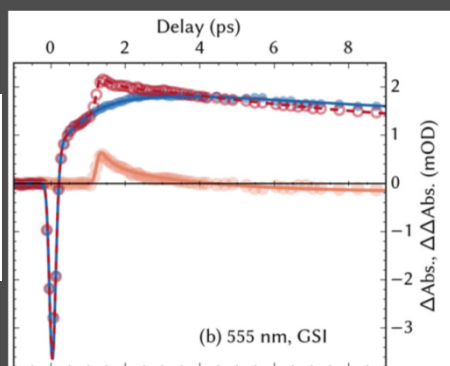
Initially, Chl a does not feel any influence of the dump.



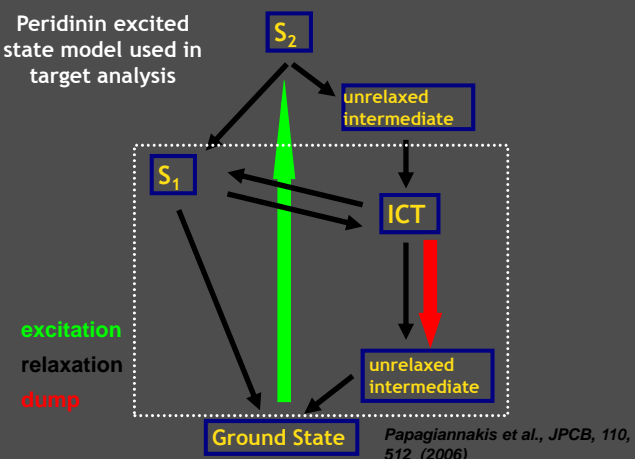
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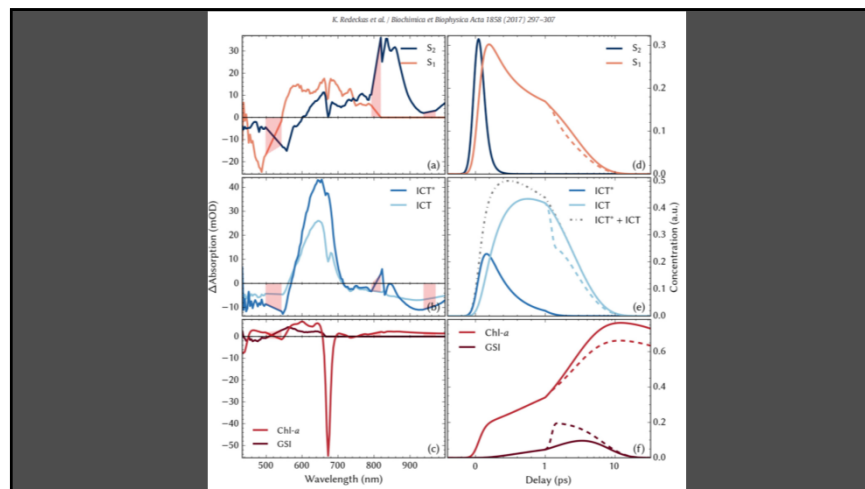
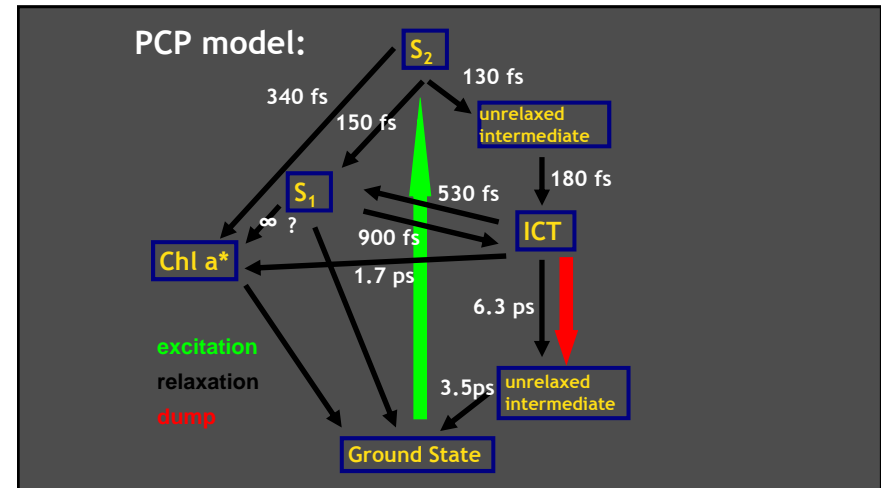
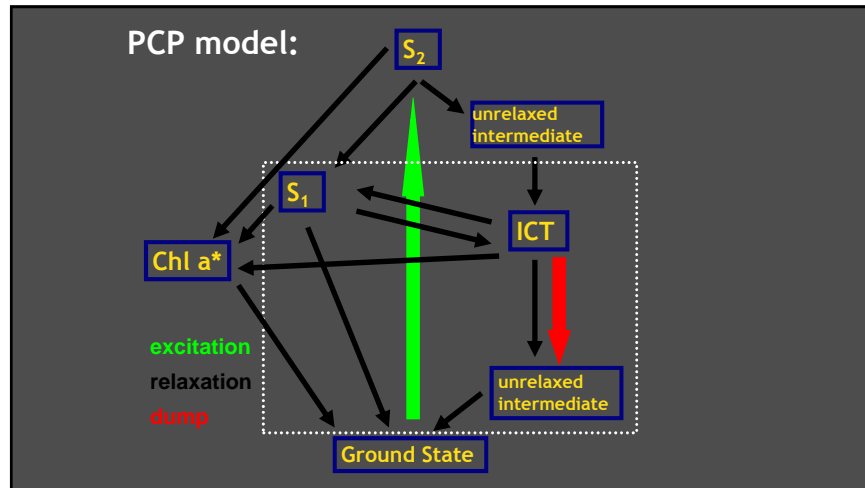


Dump produces *increase* in the IA signal – hot ground state?



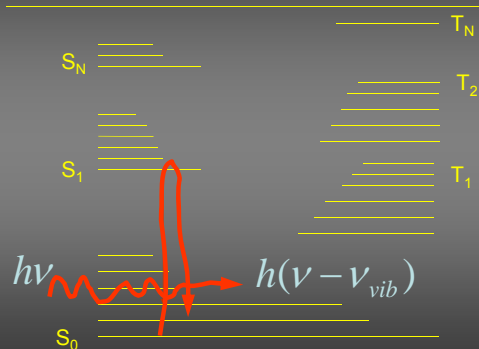
Peridinin excited state model used in target analysis





Twist #2: Femtosecond stimulated Raman spectroscopy (FSRS)

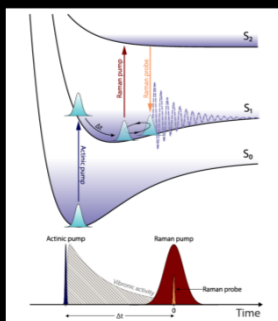
## Raman scattering



## Twist #2: Femtosecond stimulated Raman spectroscopy (FSRS)

- Playing around with electronic states is all good and well, however, the underlying structural changes are mostly guesswork.
- Raman spectroscopy is measuring vibrational frequencies, therefore it is directly sensitive to conformational changes in molecules.
- Raman spectroscopy is not prone to problems with midIR (bad detectors, ambient air absorption, etc.)
- However, to resolve narrow vibrational lines require narrow Raman Pump spectra (poor time resolution  $\sim 3$  ps).
- Enter FSRS.

## Twist #2: Femtosecond stimulated Raman spectroscopy (FSRS)

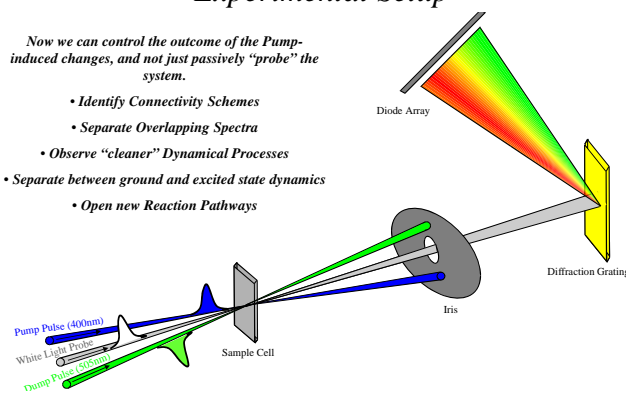


Pump-probe with a pair of probe pulses: one long (for narrow spectrum and good spectral resolution), and one short (for femtosecond time resolution).

## Dispersed Multi-Pulse Experimental Setup

Now we can control the outcome of the Pump-induced changes, and not just passively "probe" the system.

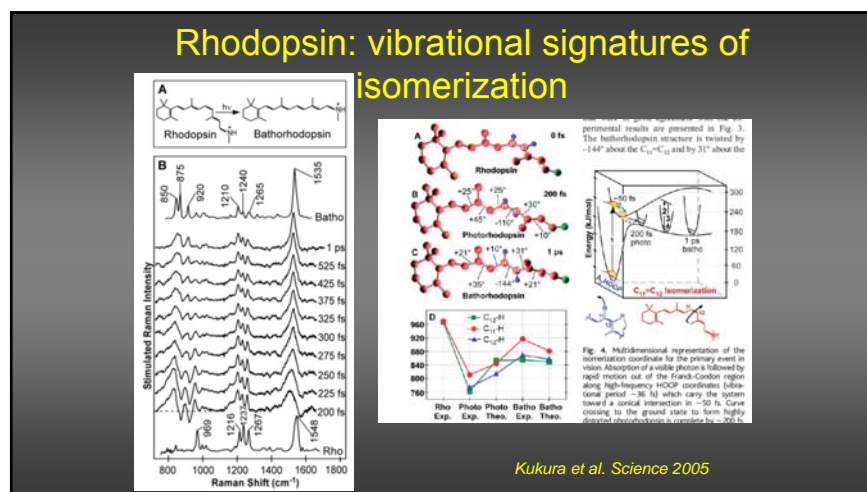
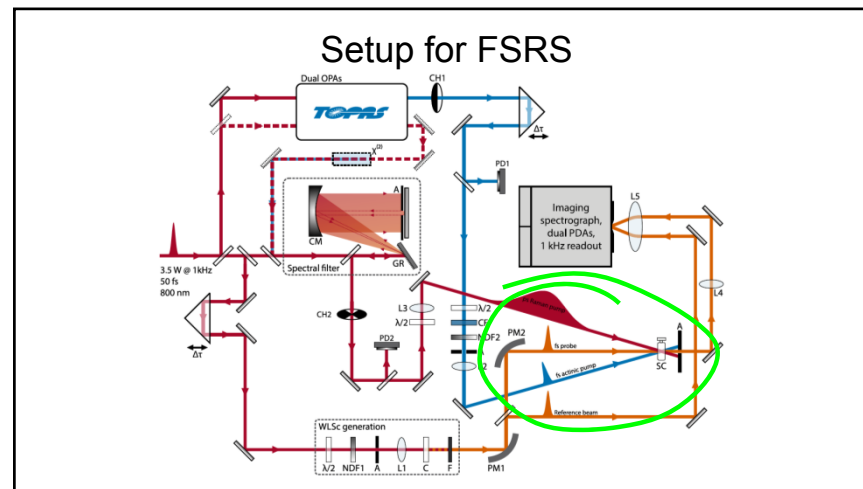
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## Dispersed Multi-Pulse Experimental Setup

*Now we can control the outcome of the Pump-induced changes, and not just passively "probe" the system.*

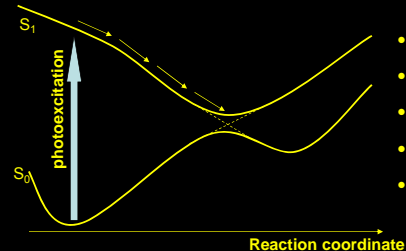
- Identify Connectivity Schemes
- Separate Overlapping Spectra
- Observe "cleaner" Dynamical Processes
- Separate between ground and excited state dynamics
- Open new Reaction Pathways



## Application: photochromism of indolo-benzoxazines

(spoiler alert: it's non-existent)

## Bistable ground-state: sensitive to everything



- Solvatochromism
- Electrochromism
- Acidochromism
- Thermochromism
- etc.

## Indolo-benzoxazines: new generation photochromic switches

Tomasulo, M.; Sortino, S.; Raymo, F. I. M. *Organic Letters* **2005**, 7, 1109.  
Tomasulo, M.; Sortino, S.; White, A. J. P.; Raymo, F. M. *Journal of Organic Chemistry* **2005**, 70, 8180. and at least 10 more papers on the same subject

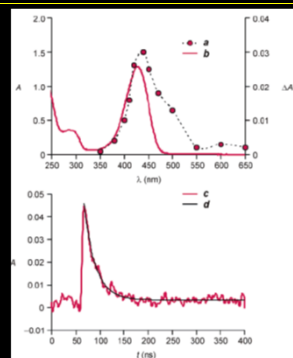
Shachkus, A. A.; Degutis, J. A.; Urbonavichyus, A. G. *Khim. Geterotsikl. Soed.* **1989**, 5, 672.



- Structure similar to spiropyrans;
- No triplet state – stable in aerobic conditions
- Fast thermal recyclization (25 ns)

## Indolo-benzoxazines: ns photodynamics

Photoinduced absorption spectrum similar to that induced by the addition of strong base ( $\text{Bu}_4\text{NOH}$ )



## Photochromism of indolo-benzoxazines

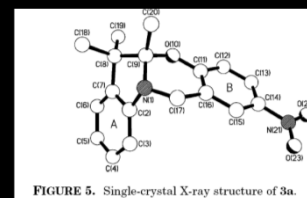
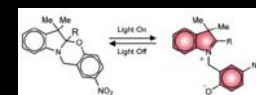


FIGURE 5. Single-crystal X-ray structure of 3a.

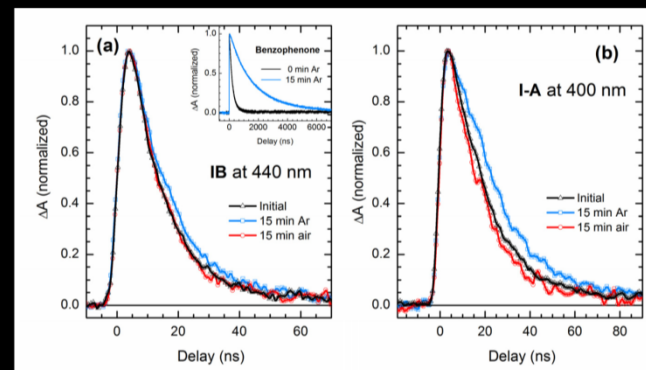


Prevailing view: UV light induces bond cleavage, and produces a p-nitrophenolate chromophore responsible for the visible absorption.

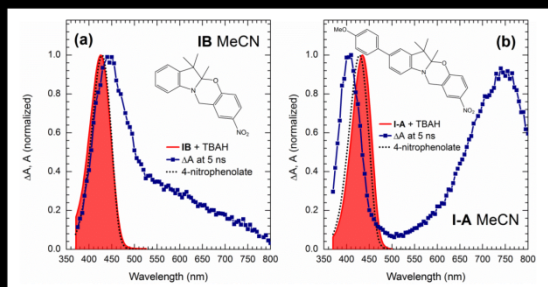


Or is it?..

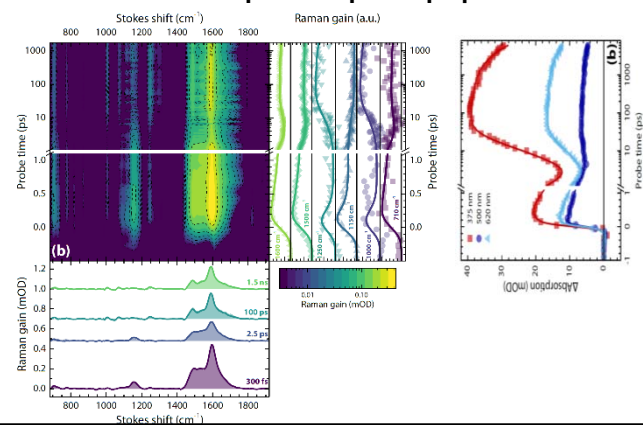
## Oxygen effect on lifetime of "isomer"



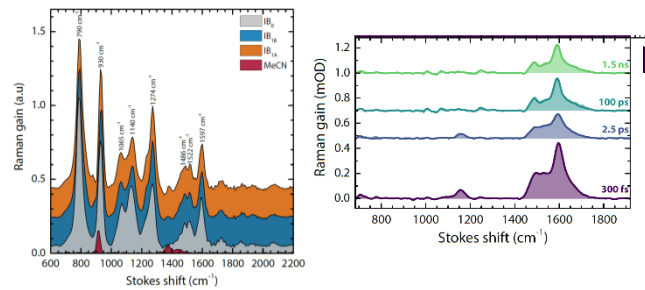
## Absorption of open and closed forms



## FSRS and optical pump-probe data



No match between optically and chemically induced Raman spectra!

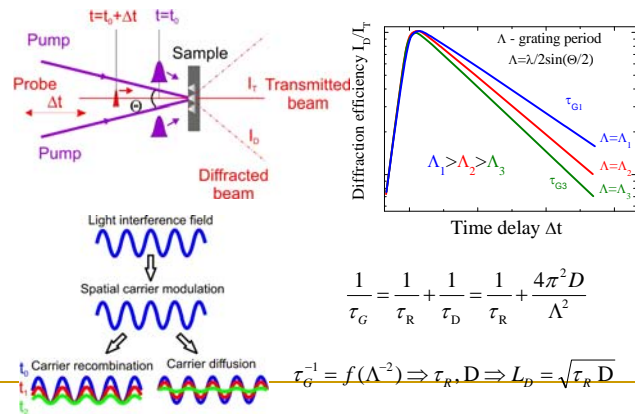


Therefore, definitely not photochromic. Pity.

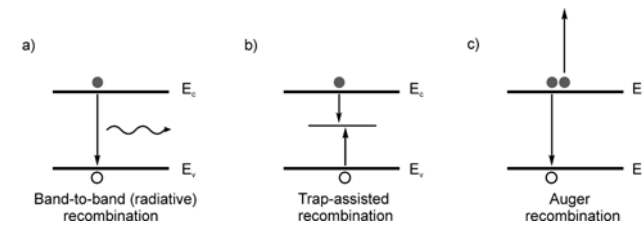
Twist #3:

Coherent spectroscopies

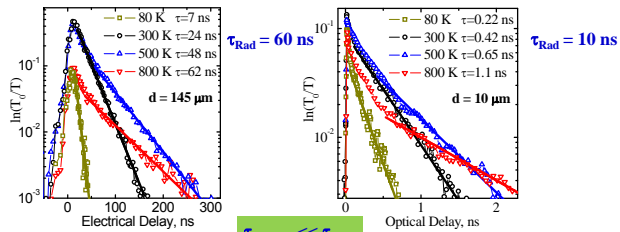
Transient grating for diffusion length determination in semiconductors



Recombination: Band-to-band (radiative), Shockley-Read-Hall (trap assisted), Auger recombination



## Non-radiative recombination time in GaN



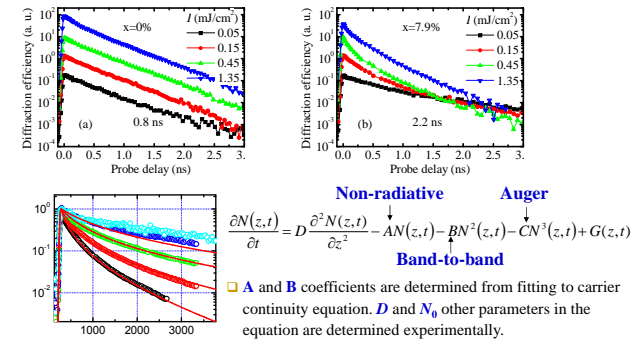
$$\frac{1}{\tau_R} = \frac{1}{\tau_{Rad}} + \frac{1}{\tau_{NonRad}}$$

$$\tau_{Rad}(T, \Delta N) = \frac{1}{B(T)\Delta N}$$

$$B(T) = 2 \times 10^{-11} \left( \frac{T}{300K} \right)^{-1.5}$$

- Non-radiative decay time  $\tau_{NonRad}$  can be determined at low injection and/or after long decay time, when  $\Delta N$  becomes small.

## Recombination in $\text{In}_x\text{Ga}_{1-x}\text{N}$ MQWs



## Transient grating

- Combined with temperature dependence, can reveal the diffusion coefficients and recombination rates and modes in semiconductors.

## More general case: three pulse echo

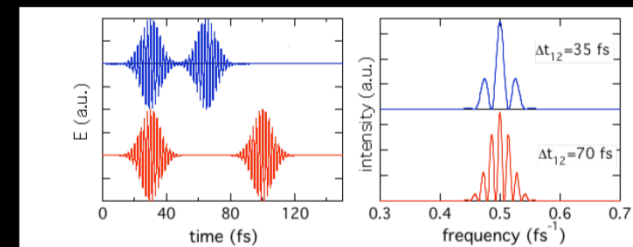
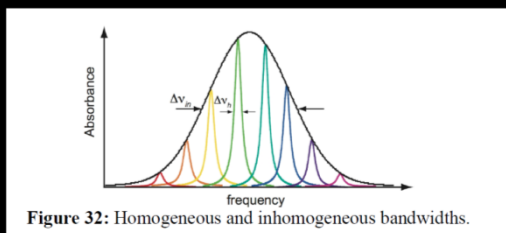
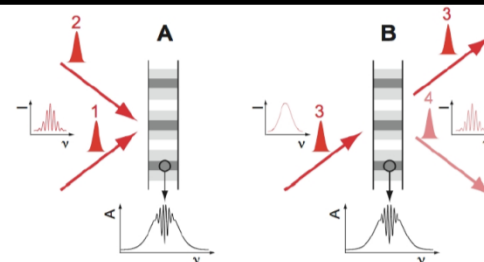


Figure 31: Electric field (left) and spectral intensity (right) associated with two 10 fs pulses separated by 35 and 70 fs.

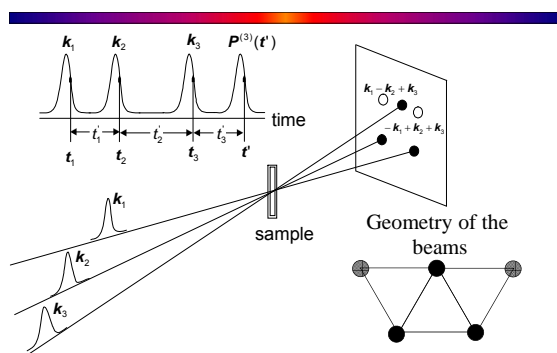
## Three pulse echo and inhomogeneous broadening



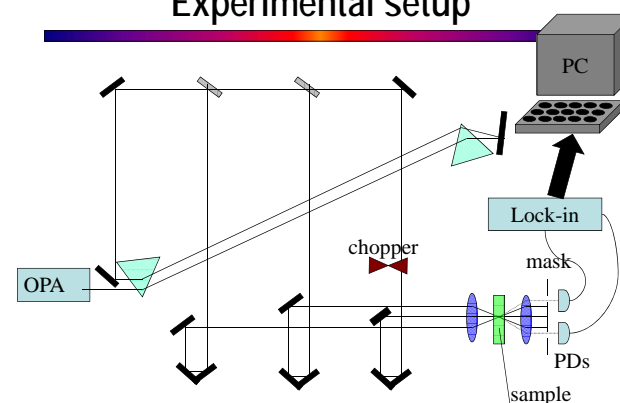
## Three pulse echo and inhomogeneous broadening



## Experiment scheme



## Experimental setup



## Three pulse echo

- The „writing“ pulses are separated in time, and their spectra are as wide as the absorption band of the sample. Shifted pulses result in frequency beating or *frequency grating*.
- The further the pulses – the finer the grating is.
- When too fine, it is very sensitive to spectral diffusion and signal quickly disappears (or, if inhomogeneous broadening is non-existent, the grating is not created to begin with;
- When too coarse, diffraction is weak
- Therefore, signal maximum is observed when the writing pulses are *SLIGHTLY* separated in time.

## Frequency gratings

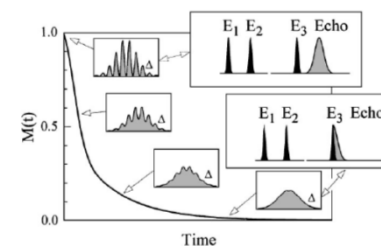
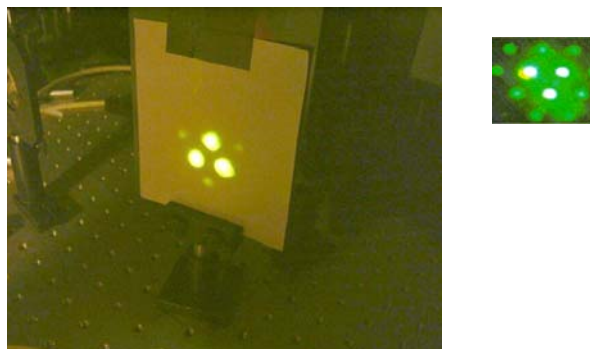


Figure 2 System-bath correlation function and associated decays of the frequency grating formed in the ground state.  $\Delta$  denotes the frequency detuning from the optical transition. The excitation pulse sequence  $E_1, E_2, E_3$  as well as the emitted signals are depicted for  $M(t) = 1$  (inhomogeneous broadening) and  $M(t) = 0$  (homogeneous broadening). Note that in the former case, the signal is delayed with respect to the last excitation pulse (photon echo), whereas in the latter case, the signal maximum coincides with the last excitation pulse (free induction decay).

## How does it look?



## What do we measure?

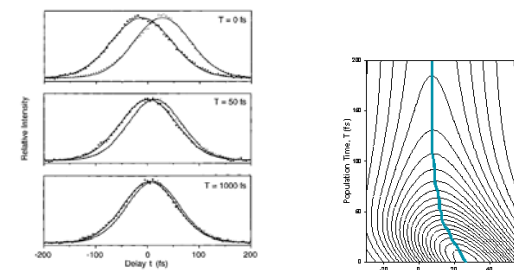
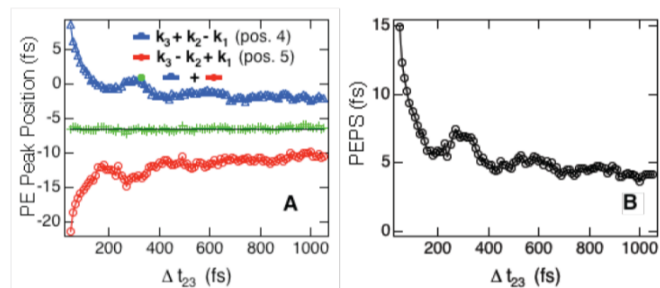


Figure 4. Examples of three 3PEPS scans (in the  $t$  dimension) obtained with  $\alpha$ -subunit preparations at room temperature at  $T = 0, 50,$  and  $1000$  fs. The two traces represent the echo signals obtained from the two phase-matched directions. The traces are superimposed with fits to Gaussian line shapes, as determined by a nonlinear least-squares regression routine.

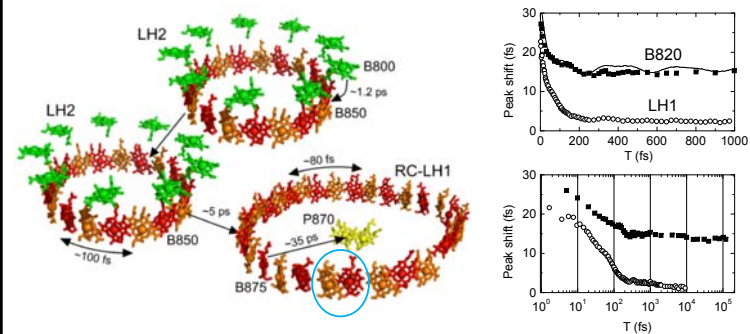
Peak shift decay follows the memory function of the bath coupled system

## Polar dye in polar solvent



**Figure 35:** Peak position of the signal intensity recorded in the 4 and 5 position (A) and PEPS (B) measured with an ethanol solution of IRI140.

## 3PEPS to monitor energy transfer



## Problems

- After all this work, just one decaying curve is measured. Cannot produce too much science with just one curve ☹.
- Data interpretation requires the microscopic model (hamiltonian) of spectral diffusion.



## 2D electronic spectroscopy

- What if we measure not just the intensity of the photon echo, but the time dependence of the radiated EM field?
- We get 2D NMR analogue in optics, called two dimensional electronic spectroscopy (2DES).

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### Phase-stabilized two-dimensional electronic spectroscopy

Tobias Brixner, Tomáš Mančal, Igor V. Stoppin, and Graham R. Fleming<sup>1)</sup>  
 Department of Chemistry, University of California, Berkeley and Physical Biosciences Division,  
 Lawrence Berkeley National Laboratory, Berkeley, California 94720

## 2D spectroscopy: plethora of pulses

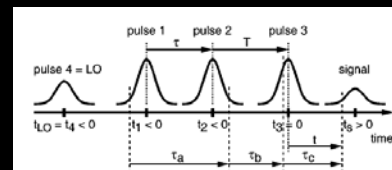


FIG. 1. Definition of time variables. Time zero is defined at the center of the third excitation pulse. The first two excitation pulses arrive at times  $t_1 < 0$  and  $t_2 < 0$ , separated by the coherence time  $\tau$  which is positive for the shown pulse order, and negative if pulse 2 arrives first. The population time  $T > 0$  is the separation between the second and third excitation pulse at  $t_3 = 0$ . Non-linear third-order polarization at time  $t$  is induced by field interactions at times  $\tau_a + \tau_b + \tau_c$ ,  $\tau_b + \tau_c$ , and  $\tau_c$  earlier, which may occur somewhere under the excitation pulse envelopes. This leads to a free-induction decay and for inhomogeneously broadened systems, an additional photon echo signal is observed with an average arrival time  $t_0$  that is similar to the coherence time. The local oscillator (LO) used for heterodyned signal detection always arrives first at time  $t_4$ .

## Experimental implementation

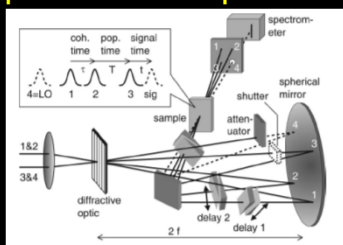


FIG. 2. Experimental setup. Two parallel beams of femtosecond laser pulses in the visible spectral region are focused by a lens onto a grating. The first diffraction orders emerge with high efficiency and provide the excitation pulses 1-3 as well as a local oscillator (4=LO) for heterodyne-detected three-pulse photon-echo electronic spectroscopy. A spherical mirror ( $2f = 50$  cm) creates an image of the pulse overlap in the sample cell via a phase folding mirror. The required time delays are provided with subwavelength precision by motor-controlled movable glass wedges. Full characterization of the nonlinear phase-matched signal field is carried out by spectral interferometry with the attenuated LO. An automated beam shutter is used for subtraction of scattering contributions. This diffractive-optics based setup is inherently phase-stabilized.

## You can calibrate your $\lambda/20$ delays

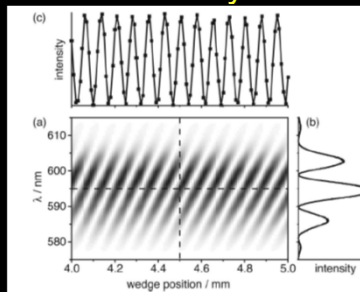
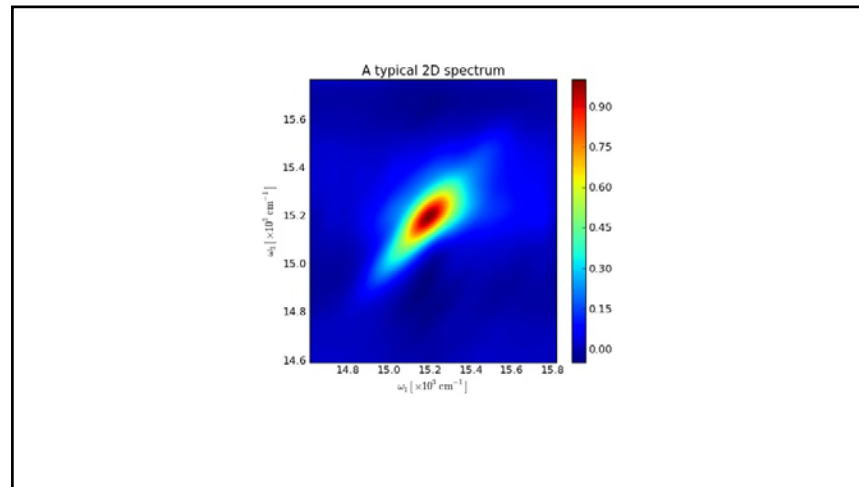


FIG. 3. Delay calibration by spectral interferometry. (a) Spectral interference patterns between pulses 1 and 2 are recorded in  $10 \mu\text{m}$  steps by moving the glass wedge in arm 1. The cross section along the vertical dashed line shows (b) the spectral interference outside of the temporal pulse overlap, whereas the cross section along the horizontal dashed line delivers (c) the temporal oscillation for one particular wavelength. Counting these oscillations gives a precise calibration factor of wedge position vs time delay.

## 2D Electronic spectroscopy

- Can distinguish homogeneous and inhomogeneous broadening;
- Time resolution is not limited by spectral resolution on excitation scale;
- It's like many pump-probe experiments in one go!



## 2D electronic spectroscopy

The multidimensional Fourier transform of the 2D spectrum is given by the 2D correlation function  $C^{(2)}(\omega_1, \omega_2, \tau)$ . The 2D correlation function is defined as the Fourier transform of the 2D spectrum  $S^{(2)}(\omega_1, \omega_2, \tau)$  with respect to the detection energy  $\omega_2$  and the time delay  $\tau$ . The 2D correlation function is a function of the excitation energy  $\omega_1$ , the detection energy  $\omega_2$ , and the time delay  $\tau$ . The 2D correlation function is a function of the excitation energy  $\omega_1$ , the detection energy  $\omega_2$ , and the time delay  $\tau$ . The 2D correlation function is a function of the excitation energy  $\omega_1$ , the detection energy  $\omega_2$ , and the time delay  $\tau$ .

- Requires detailed QM theory
- High coolness factor

## 2D electronic spectroscopy: what do we measure?

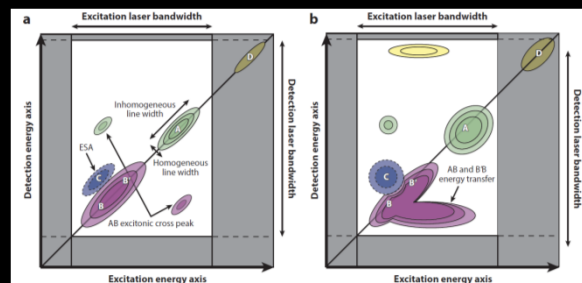
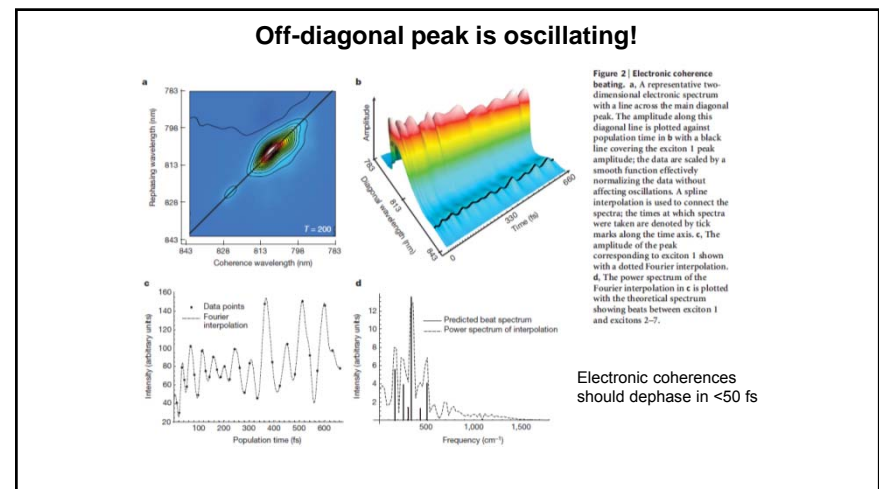
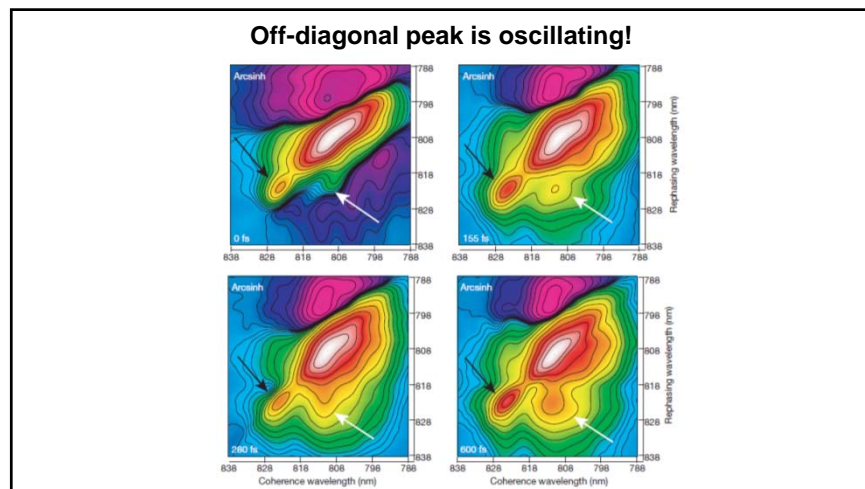
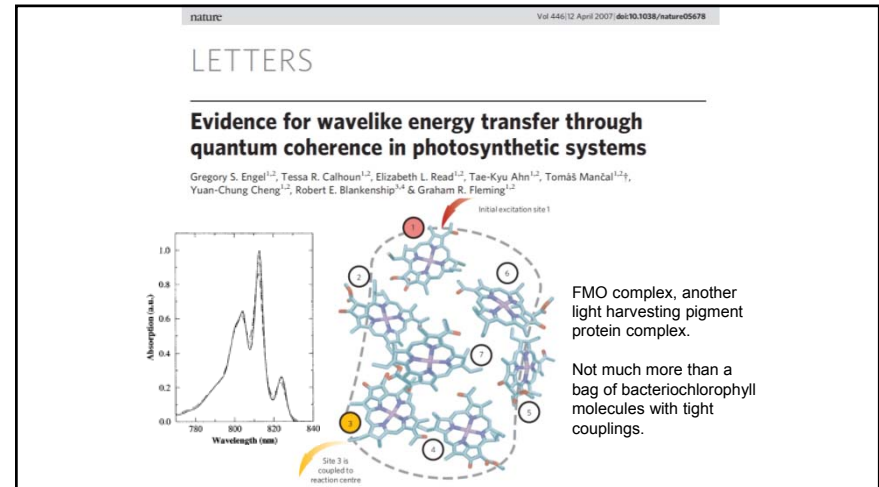


Figure 2 Schematic illustration of the information contained in absorptive two-dimensional Fourier transform spectra. The observed spectral range shown in the unshaded region is determined by the excitation and detection laser bandwidths. (a) The  $T = 0$  correlation spectrum reveals homogeneous and inhomogeneous line widths, excitonic coupling, and excited state absorption features. (b) At  $T > 0$ , the broadening of peaks in the anti-diagonal direction reflects spectral diffusion. The growth of cross peaks indicates energy transfer. The emergence of an entirely new peak at later waiting times, such as the B'D cross peak in panel (b), represents the formation of a new product species, populated upon excitation of B', that absorbs at D. An example of such a process is the generation of a charge-separated state in the photosystem II reaction center (data shown in Figure 4).



If you are from Berkeley, you can  
publish anything you like!



This 'wavelike' energy transfer started the now fashionable (hopefully, not for long) field of quantum biology...

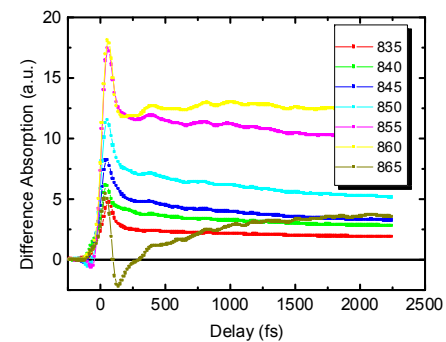
My never published data from 2000

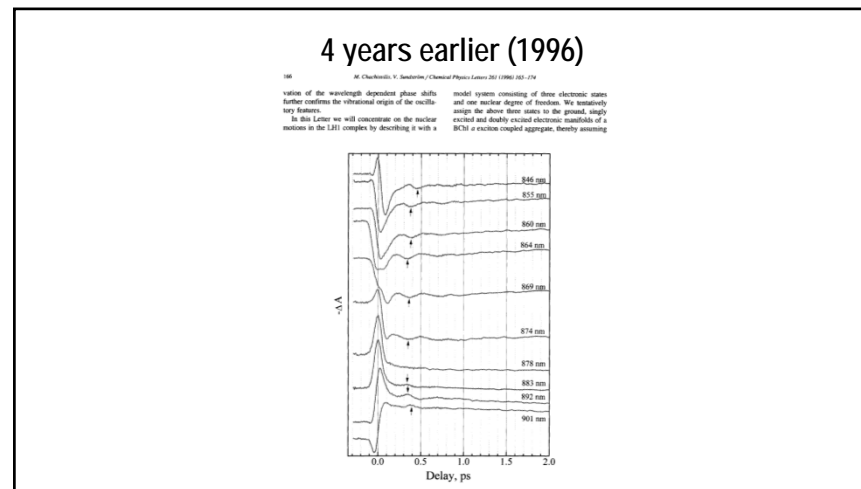
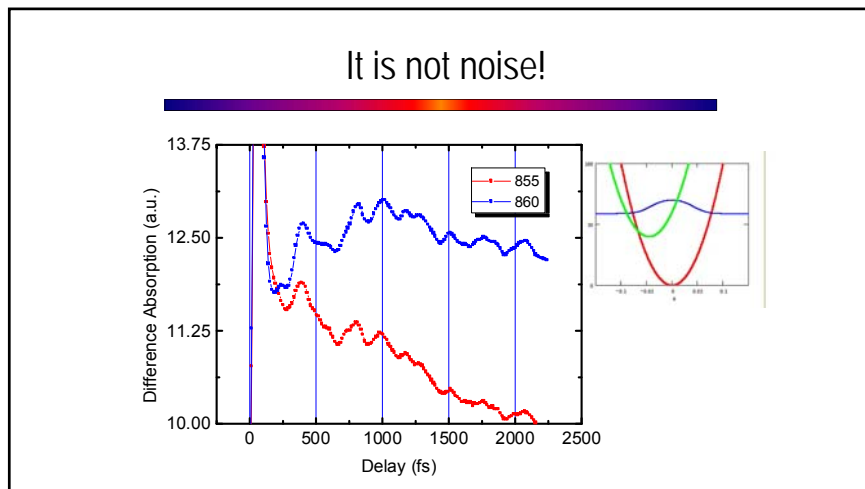
## Coherent response of nucleic subsystem to the excitation.

- Good old Rps. Acidophila;
- Pump-probe at 77K;
- Purpose: observe how the nuclei of BChl molecules respond to the electronic excitation.



## Kinetics at various wavelengths (77K)





## Instead of conclusions...

Time-resolved spectroscopy comes in a lot of different guises and is used in different fields for understanding the quantum-mechanical functioning of light sensitive matter.

Used wisely, it is a powerful box of tools for investigating nature.

Used bravely, it gets you papers in Nature (sometimes even despite the science being wrong).