



Determination of the protein-ligand binding volume by high-pressure spectrofluorimetry

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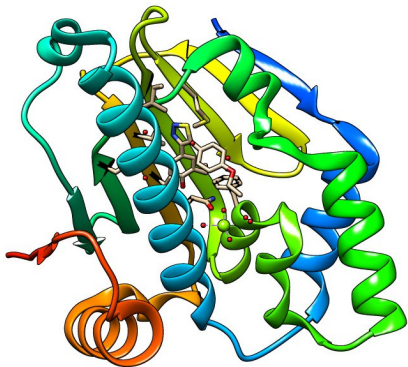
Introductory remarks

Protein-ligand binding (reaction) volume

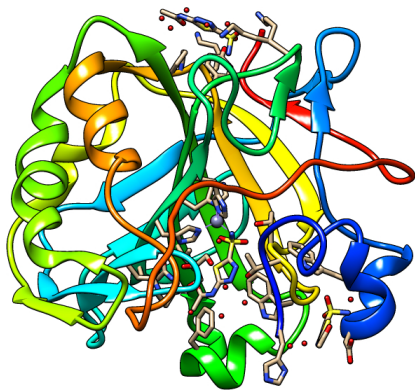
- Definition: the change in protein volume observed upon protein-ligand binding;
- Motivation: fundamental knowledge about protein-ligand interaction and pressure-induced protein denaturation.
- Protein-ligand binding volume: $\Delta V_b = \left(\frac{\partial \Delta G_b}{\partial P} \right)_T$;

Introductory remarks

Heat shock protein 90 and carbonic anhydrases

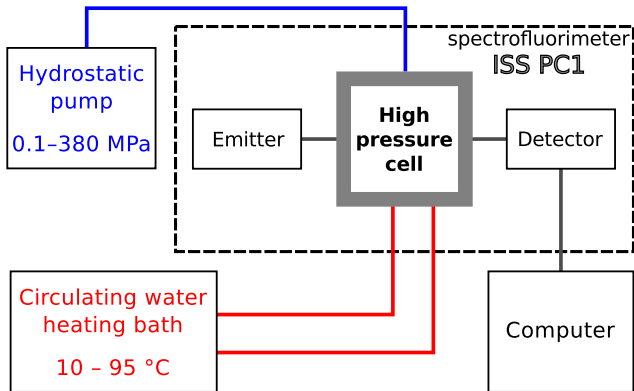


N-terminal domain of human Hsp90
(229 a.a.), PDB ID: 2YI6



Human carbonic anhydrase (CA) II
(260 a.a.), PDB ID: 3HS4

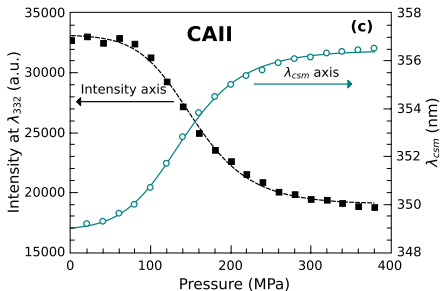
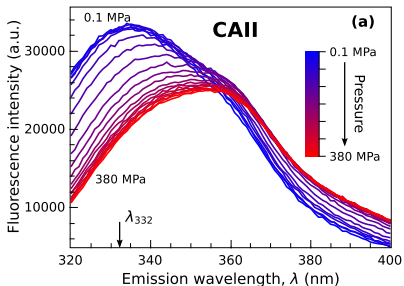
Experimental set-up of high-pressure spectrofluorimetry



- Pressure range: (0.1 – 380) MPa;
- Temperature range: (10 – 95) °C;
- Fluorescence probe: intrinsic tryptophan (excited at 295 nm);
- Emission spectra range: (320 – 400) nm.

Description of pressure-induced unfolding profiles

Equations



Unfolding profiles:

$$f(p) = f_N + \frac{f_U - f_N}{1 + \exp(\Delta G(p)/RT)}$$

The Gibbs energy of unfolding:

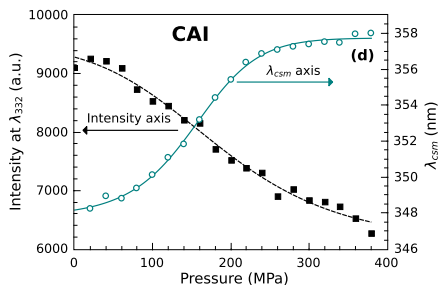
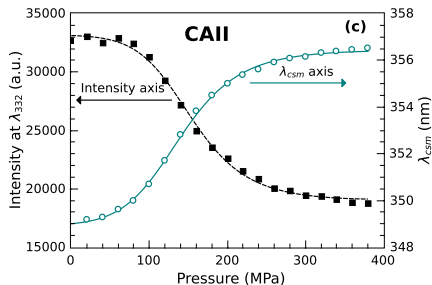
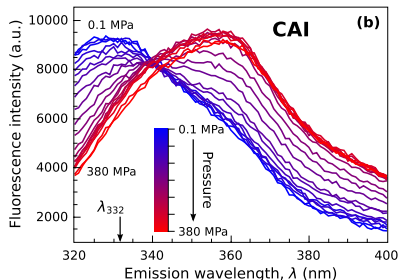
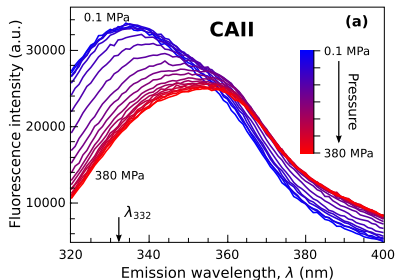
$$\Delta G = \Delta G_0 + \Delta V_0 \Delta p + \frac{\Delta \beta}{2} (\Delta p)^2$$

The center of spectral mass:

$$\lambda_{CSM} = \frac{\sum_i f_i \lambda_i}{\sum_i f_i}$$

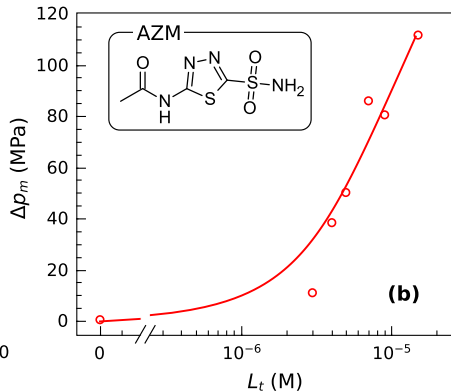
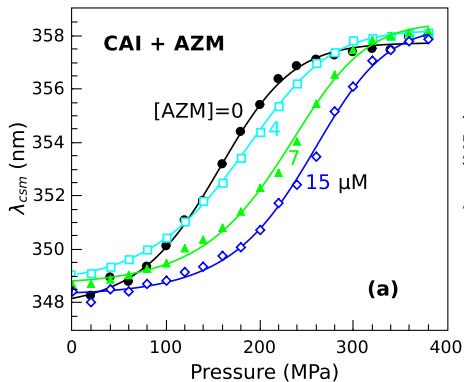
Pressure-induced unfolding profiles of CA II and CA I

Intrinsic tryptophan fluorescence spectra at various pressures



The shift in P_m and dosing curves

Fluorescent Pressure Shift Assay (FPSA)



Description of dosing curves

Mathematical background

$$P_t = N_f + N_b + U,$$

$$L_t = L_f + N_b,$$

$$K_U = \frac{U}{N_f},$$

$$K_b = \frac{N_b}{N_f L_f}.$$

P_t and L_t – the bulk concentrations of protein and ligand;

N_f and N_b – concentrations of native ligand-free and bound protein;

U – concentration of unfolded protein;

K_U and K_b – equilibrium constants of protein unfolding and protein-ligand binding reaction;

At melting pressure p_m , $U = P_t/2$.

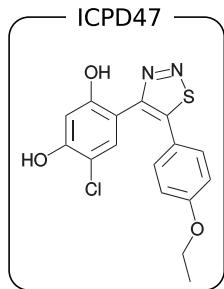
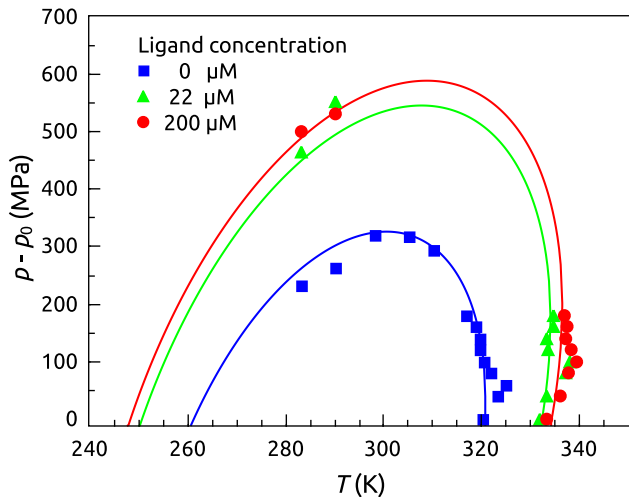
The final form of dosing curve equation:

$$L_t = (K_U - 1) \left(\frac{1}{K_b} + \frac{P_t}{2K_U} \right)$$

$$K_b = \exp \left(-\frac{\Delta G_b}{RT} \right) = \left(-\frac{\Delta G_{0_b} + \Delta V_{0_b}(p_m - p_0) + \frac{\Delta \beta_b}{2}(p_m - p_0)^2}{RT} \right)$$

Hsp90N protein stability diagram in P - T coordinates

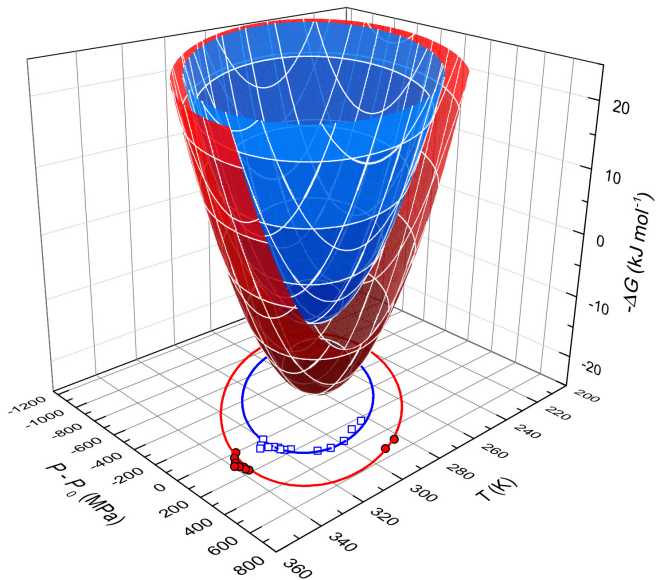
With added ligand



 Petrauskas et al. *Eur Biophys J* 42 (2013) 355–362.

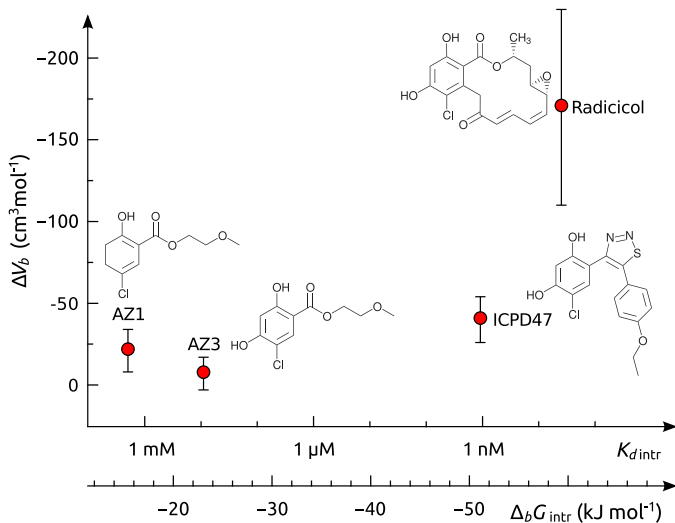
Hsp90N protein stability diagram

The Gibbs energy dependence on pressure and temperature



Binding volume correlation with affinity

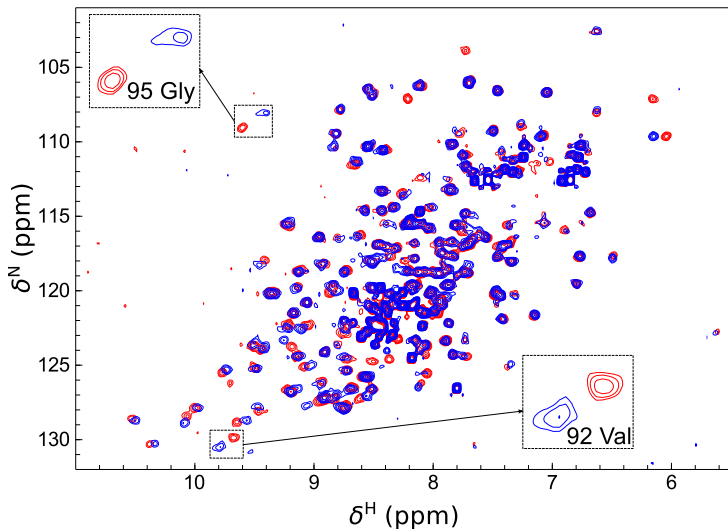
Affinity was determined by ITC and Fluorescent Thermal Shift Assay



- Preliminary results of Hsp90N interaction with two ligands by NMR;
- $\Delta V_b = \left(\frac{\partial \Delta G_b}{\partial p} \right)_T$;

NMR data

^1H - ^{15}N -HSQC spectra of Hsp90N (red) and Hsp90N+ICPD9 (blue)



NMR data analysis

Analysis of dosing curves

$$\Delta V_b = \left(\frac{\partial \Delta G_b}{\partial p} \right)_T$$

$$\Delta G_b = -RT \ln(K_b) = RT \ln(K_d)$$

Experimental changes in chemical shifts:

$$\Delta\delta = \sqrt{(\delta_0^H - \delta^H)^2 + \left(\frac{\gamma^N}{\gamma^H} \right)^2 (\delta_0^N - \delta^N)^2}$$

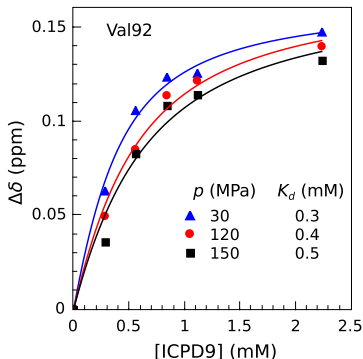
δ_0 – chemical shift without ligand.

$$\Delta\delta = \Delta\delta_{max} \frac{(L_t + P_t + K_d) - \sqrt{(L_t + P_t + K_d)^2 - 4P_t L_t}}{2P_t}$$

Chemical shift equation reference:

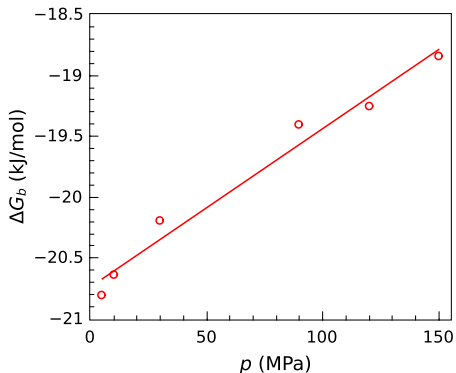


Morton et al. *Structure* 4 (1996) 705-14.



Gibbs energy as a function of pressure

$$\Delta G_b = \Delta G_{b_0} + \Delta V_{b_0} \Delta p + \frac{\Delta \beta_b}{2} (\Delta p)^2.$$



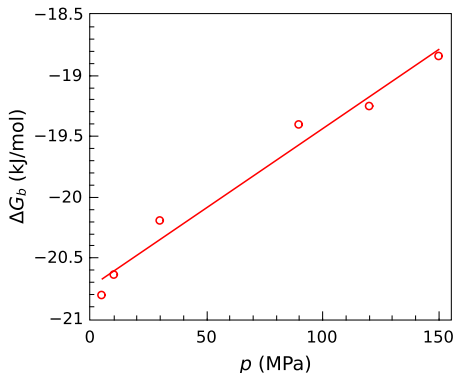
$$\Delta G_{b_0} = -20.7 \text{ kJ/mol}$$

$$\Delta V_{b_0} = 13 \text{ cm}^3/\text{mol}$$

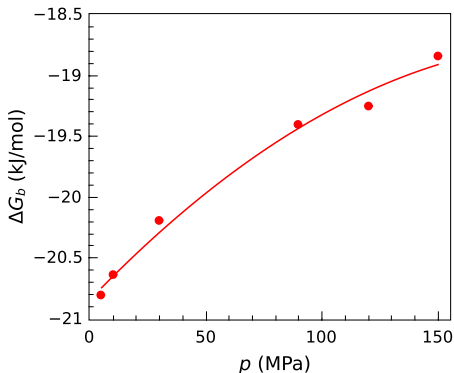
$$\Delta \beta_b = 0 \text{ cm}^6/(\text{J mol})$$

Gibbs energy as a function of pressure

$$\Delta G_b = \Delta G_{b_0} + \Delta V_{b_0} \Delta p + \frac{\Delta \beta_b}{2} (\Delta p)^2.$$



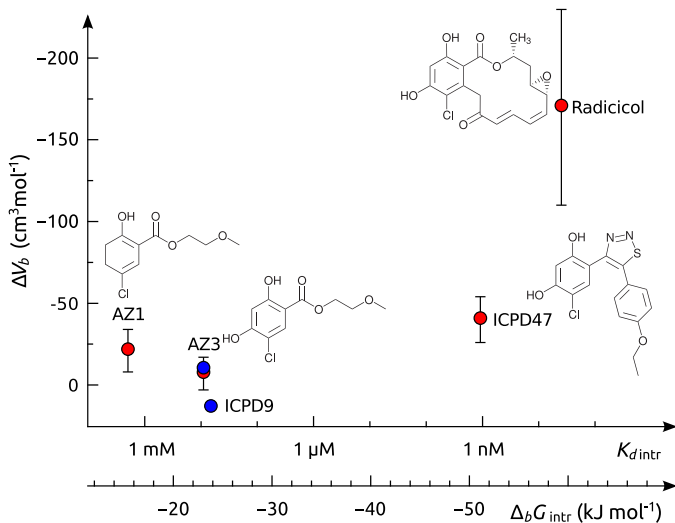
$$\begin{aligned}\Delta G_{b_0} &= -20.7 \text{ kJ/mol} \\ \Delta V_{b_0} &= 13 \text{ cm}^3/\text{mol} \\ \Delta \beta_b &= 0 \text{ cm}^6/(\text{J mol})\end{aligned}$$



$$\begin{aligned}\Delta G_{b_0} &= -20.8 \text{ kJ/mol} \\ \Delta V_{b_0} &= 20 \text{ cm}^3/\text{mol} \\ \Delta \beta_b &= 0.05 \text{ cm}^6/(\text{J mol})\end{aligned}$$

Binding volume correlation with affinity

Including NMR data (blue points)



- Our results suggest that weak ligands of Hsp90N protein induce smaller changes in volume than tight-binding ligands.
- Both techniques FPSA and NMR provide similar values of binding volume.

Acknowledgments

High pressure team at the department of
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NMR data:

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Lietuvos
mokslo
taryba



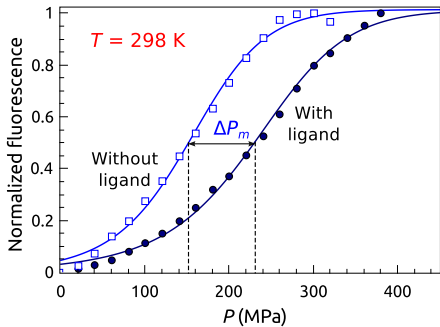
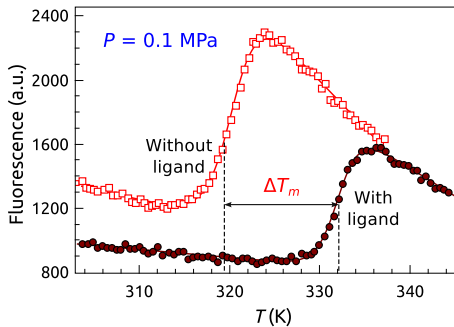
High-pressure team

Thank you for your attention!

Supplementary material

FTSA and FPSA methods

Examples of Hsp90N protein stabilization by ligand against T and P denaturation



FTSA and FPSA methods

Expressions for the Gibbs energy

$$f = f_N + \frac{f_U - f_N}{1 + \exp(\Delta_U G / RT)}$$

$\Delta_U G$ as a function of **temperature** at a constant **pressure**:

$$\Delta_U G_T = \Delta_U G_0 - \Delta_U C_p \left[T \left(\ln \frac{T}{T_0} - 1 \right) + T_0 \right] - (\Delta_U H_0 - \Delta_U G_0) \left(\frac{T}{T_0} - 1 \right)$$

$\Delta_U G$ as a function of **pressure** at a constant **temperature**:

$$\Delta_U G_P = \Delta_U G_0 + \Delta_U V_0 (p - p_0) + \frac{\Delta_U \beta}{2} (p - p_0)^2$$

The Gibbs energy change

The full differential of the Gibbs energy change $\Delta G = G_U - G_N$

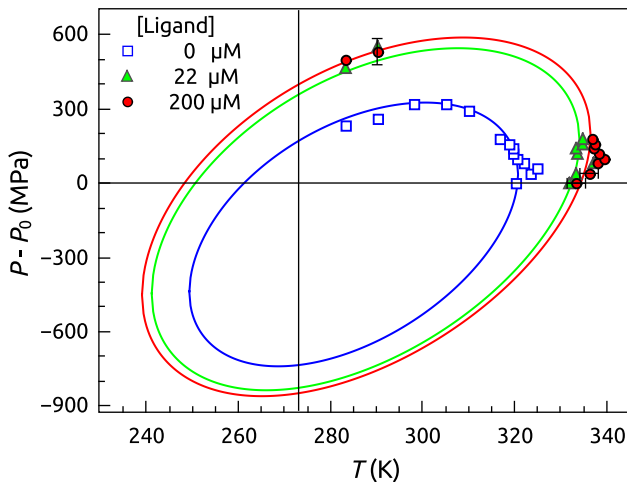
$$d(\Delta G) = -\Delta S dT + \Delta V dp$$

as a function of temperature T and pressure P .

$$\Delta G = \frac{\Delta\beta}{2} (p - p_0)^2 + \Delta V_0(p - p_0) + \Delta\alpha(p - p_0)(T - T_0) - \Delta C_p \left[T \left(\ln \frac{T}{T_0} - 1 \right) + T_0 \right] - (\Delta H_0 - \Delta G_0) \left(\frac{T}{T_0} - 1 \right) + \Delta G_0$$

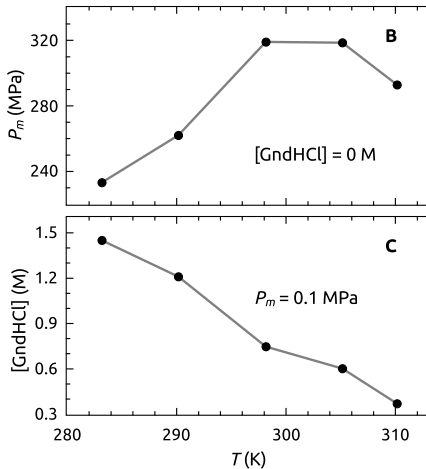
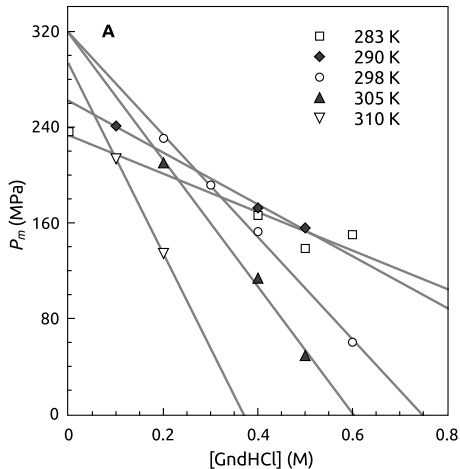
Hsp90N protein stability diagram in P - T coordinates

With added ligand



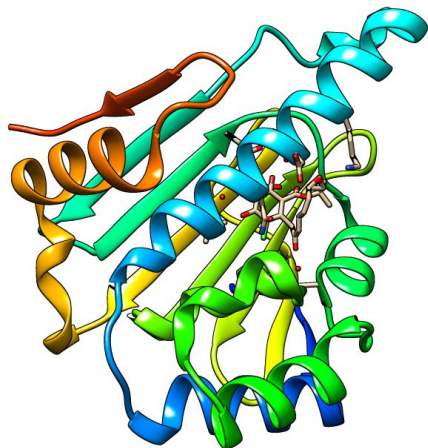
Hsp90N stabilization by ligands

The use of guanidine hydrochloride – a protein destabilizing agent



Protein stabilization by ligands

Hsp90N + radicicol



Hsp90N + radicicol
PDB ID: 4EGK

Hsp90N melting temperatures T_m

$[\text{Hsp90N}] = 14 \mu\text{M}$

$[\text{Radicicol}] \mu\text{M}$	T_m ($^{\circ}\text{C}$)
0	51.6
2	61.1
20	64.9



Zubriene et al. *Int J Mol Sci* **10** (2009) 2662–2680.